

Electronic-nose Applications in Forensic Science and for Analysis of Volatile Biomarkers in the Human Breath

Wilson AD*

Forest Insect and Disease Research (FIDR), Southern Hardwoods Laboratory, Stoneville

*Corresponding author: Wilson, AD. Ph.D., Principal Research Pathologist, USDA Forest Service, Southern Hardwoods Laboratory, Stoneville, MS 38776-0227, Tel: +1 662 6863180, Fax: +1 662 6863195, E-mail: dwilson02@fs.fed.us

Citation: Wilson AD (2014) Electronic-nose Applications in Forensic Science and for Analysis of Volatile Biomarkers in the Human Breath. *J Forensic Sci Criminol* 1: S103. doi: 10.15744/2348-9804.1.S103

Received Date: May 14, 2014 **Accepted Date:** June 06, 2014 **Published Date:** June 26, 2014

Abstract

The application of electronic-nose (E-nose) technologies in forensic science is a recent new development following a long history of progress in the development of diverse applications in the related biomedical and pharmaceutical fields. Data from forensic analyses must satisfy the needs and requirements of both the scientific and legal communities. The type of data collected from electronic-nose devices provides a means of identifying specific types of information about the chemical nature of evidentiary objects and samples under investigation using aroma signature profiles of complex gaseous mixtures containing volatile organic compounds (VOCs) released from manufactured products and parts of the human body. E-nose analyses also provide useful qualitative information about the physicochemical characteristics and metabolic conditions of human subjects without the need for time-consuming analyses to identify all chemical components in human-derived volatile mixtures. E-nose devices are capable of providing information for a wide range of forensic applications, useful for answering many types of questions relating to past events and details of circumstances and conditions that led to criminal activities involving human subjects and the perpetrators involved. E-nose devices have been used to help locate live subjects, buried in the rubble of collapsed buildings following natural disasters, as well as hidden bodies and the human remains of victims of accidents and crimes of aggression. The noninvasive analysis of gaseous mixtures in the human breath and lungs of living and deceased individuals provides a means for identifying the existence of diseases or adverse physiological conditions of human subjects (both before death and postmortem) potentially useful in determining the cause of death, time of death, and pertinent factors contributing to lethal events such as homicides and other violent crimes.

Keywords: Artificial olfaction; Biomarker indicator compounds; Breath gas analysis; Cadaverine; Disease diagnostics; Electronic aroma detection; E-nose; Metabolomics; Respiratory gas metabolites; Volatile organic compounds

Introduction

The continuous improvement in methods and tools used to facilitate the acquisition of evidence gathered in criminal, forensic and cause-of-death investigations requires the recognition and implementation of new technologies that provide either new types of information, corroborative evidence, or more detailed information by more accurate, rapid or efficient means. The development and use of new forensic analytical technologies ultimately expedite the progress of criminal investigations, leading to more rapid and conclusive resolutions of judicial processes through litigations. Many new forensic tools for chemical analyses have been developed over the years to provide more effective analyses of different sample types. Electronic-nose (E-nose) instruments represent new types of electronic aroma detection (EAD) technologies that are being developed for numerous applications in the fields of forensics and criminology [1,2], and related biomedical and pharmaceutical industries [3,4]. There are many different types of E-nose devices including surface acoustic wave (SAW), quartz crystal microbalance (QMB), metal oxide semiconducting (MOS), conducting polymers (CP), and others [5], as well as the more recent carbon nanotube types (see paper by Kybert et al. in this same issue-Sniffing Out Human Odor).

Electronic-nose devices generally are used primarily to detect and identify specific gaseous mixtures of volatile chemical compounds, including organic and inorganic chemicals released from material sources, rather than identify individual chemical compounds present in sample mixtures. Thus, the sensor output from an E-nose analysis of a sample reflects the combined aroma characteristics of all the chemical constituents present in the sample as a whole. This information is different from most conventional forensic chemical analyses required to determine the precise chemical composition of evidentiary samples involved in criminal cases. However, E-nose instruments are capable of identifying individual organic and inorganic compounds present in pure form or in simple gaseous mixtures when trained to do so. An E-nose device identifies specific gaseous mixtures or individual compounds in the sample by comparing the output from the E-nose sensor array to reference databases, produced by instrument-training to recognize known mixtures or compounds, based on mathematical and statistical processes involving pattern recognition algorithms [5].

This review provides a synopsis of some potential E-nose applications available for chemical analyses in the fields of forensic science, criminology, and medical diagnostics (autopsies etc.) that complement conventional chemical methods used to analyze different forensic sample types. The remainder of this review focuses on specific examples of electronic-nose applications for the detection and analysis of volatile metabolites in the human breath, particularly biomarkers (respiratory metabolites and other respired chemicals), that serve as indicators of specific causes of human ailments, including diseases and metabolic disorders, that contribute to information pertinent to the causes of natural fatalities or deaths associated with various types of forensic investigations.

I Potential E-nose applications in forensics

Electronic-nose instruments recognize precise gaseous mixtures of volatile organic compounds (VOCs) by the unique “fingerprint” pattern or sensor profiles resulting from sensor responses to VOC gases generated from the collective output of cross-reactive sensors in an E-nose sensor array. The combined output pattern (aroma profile) from the multisensor array is produced in response to all of the VOCs present in the sample mixture as these compounds are adsorbed and detected by individual sensors in the sensor array. Different types of electronic-nose instruments utilize different mechanisms for detection although most E-nose detection systems include a transducer that converts the electronic detection signal from sensors into digital output values to record individual sensor responses that makeup the combined aroma output pattern. The many and varied types of E-nose instruments available for chemical analyses have been summarized previously [5].

The types or chemical classes of VOCs detected in forensic, criminology, and diagnostic investigations vary widely depending not only on the many different forensic sample types being chemically analyzed, such as human manufactured products and human body-derived samples (tissues, fluids, and exhaled gases), but also the purpose or investigative intent of the chemical analysis. E-nose instruments, utilizing different chemical-detection mechanisms and technologies, are capable of sensing a wide range of volatile inorganic compounds (VICs) and VOCs from a large diversity of chemical classes. A wide range of E-nose instrument types are available for EAD analyses of samples from many different types of forensic and criminal investigations. The chemical constituents present in evidentiary samples, range from VOCs released from manmade products to respiratory metabolites from living human patients and microbial degradation products released from dead human remains. Some of the major chemical classes of VICs and VOCs analyzed in various types of forensic chemical analyses (based on sample types) are listed in Table 1 along with conventional analytical methods used for identification and some potential corresponding E-nose methods available for detections and identifications of each sample type.

Sample types ¹	Categories	Chemical classes ¹	Example compounds ²	Conventional analyses ¹	Conv. Refs.	E-nose Refs. ³
Arson	Accelerants (ignitable liquids)	HC fuels	petrol (gasoline), kerosene, paint thinners	PVD, GC, GC-MS, GC-IRMS	[6,7]	[8-11]
	Fiber dyes	triazines	dichlorotriazine	TLC, HPLC, SERRS, RR, LC-MS	[6,12-14]	NR
CTE	Paint pigments, extenders, binders	organic/inorganic particulates	Volatile paints/solvents, non-volatile particulates	PLM, FM, FTIR, XRD, XRF, SEM-EDS, LA-ICP-MS, PGC, PMS	[6,15]	[16,17]
	Glass	inorganics	Non-volatile inorganics	GRIM, SEM-EDS, XRF, ICP-AES, ICP-MS, LA-ICP-MS, SEM,	[6,18,19]	NR
	Soil	inorganics, organics	non-volatile inorganics, various VOCs	ICP-AES, ICP-MS	[6,20]	[17,21]
	Cosmetics	liquids and solids emitting VOCs	face powder, lipstick, mascara, eye liner, nail polish, perfumes, lotions	XRD, XRF, SEM/EDS, SERRS	[6,13]	[22,23]
	Shoe polish	various HCs	waxes, pigments, aniline dyes; nitrobenzene	XRD, XRF, SERRS	[6,13]	[24]
	Documents	Inks, solvents	various HCs	Ink pigments, 2-phenoxyethanol	VLMS, TLC, RM, SERRS, LC-MS	[6,13,25-28]
Paper		VOCs	Volatile byproduct residues in manufactured paper	FTIR	[31]	[32]
Explosives	Primary	inorganics	Lead azide, tetrazene, mercury fulminate	XRF	[6]	[17]
	Secondary	aromatic HC	nitrocellulose, HMX, TNT, TATB, PETN, RDX, picric acid, tetryl	IMS, HPLC, MS, GC-TEA, LC-MS	[33-36]	[37-41]
	Propellants	organics	black powder (potassium nitrate, charcoal, sulfur), nitrocellulose	HPLC, GC-TEA, LC-MS, XRF	[6,36,42]	[43,44]

FDRs	Primers	inorganics	lead styphnate, antimony sulfide, barium nitrate; also Zn- and Ti-containing particles	SEM-EDS,	[45]	[17]
	Propellants	organics	nitrocellulose, nitroglycerine, nitroguanidine	HPLC, GC-TEA, LC-MS, XRF	[25,45]	[46-48]
Human body fluids	Blood, excrement, oral fluid, semen, sweat, urine, sputum, etc.	DNA, VOCs, inorganics	nucleic acids, complex VOCs profile, inorganic contaminants	DNA profiling, XRF	[6]	[3,4]
NFDRs	Pyrotechnics, fireworks, automobile brake pads	inorganics	lead styphnate, antimony sulfide, barium nitrate	XRF	[45,49]	[17]
Toxins	Alcohol	aliphatic alcohol	ethanol	SFST, DRE, FIT	[6,18]	[17,50-52]
	Drugs	phenethylamines	amphetamine, MDA, MDMA, epinephrine, norepinephrine, Dopamine	MT, TLC, GC-MS, NMR, HPLC, EIA, RM	[53-55]	[3,5]
		barbiturates	Phenobarbital, Barbitol, Alphenal	LC-MS, HPLC, GC-MS, IRS	[33]	[3]
		benzodiazepines	Diazepam, Lorazepam	GC-MS, HPLC-uv	[6,33]	NR
		opioids	Morphine, Fentanyl, Etorphine, Heroin, Methadone (synthetic)	XRD, MT, HPLC, GC-MS, EIA	[6,33,37,53-55]	NR
		tropane alkaloids	Cocaine, Ecgonine, Norcocaine, Benzoylcegonine, Truxillines	CIT, ST, TLC, GC-MS, SIRMS, EIA, NMR	[6,33,37,56-58]	[59]
		cannabinoids	Cannabis (Δ^9 -THC)	TLC, EIA, GC-MS, HPLC	[6]	[60]
Heavy metal poisoning	inorganics	Arsenic, cadmium, lead, mercury, osmium, thallium, vanadium	XRF	[61]	[17]	

¹**Abbreviations:** CIT = Cobalt Isothiocyanate Test; CTE = Chemical Trace Evidence; DRE = Drug Recognition Examinations; EIA = Enzyme Immunoassay; FDRs = Firearm discharge residues; FIT = Field Impairment Testing; FM = Florescence Microscopy; FTIR = Fourier Transform Infrared Spectroscopy; GC-MS = Gas Chromatography with Mass Spectrometry; GC-TEA = Gas Chromatography with Thermal Energy Analysis; GRIM = Glass Refractive Index Measurement; HC = Hydrocarbons (organic compounds containing carbon and hydrogen); HPLC = High Performance Liquid Chromatography with amperometric detection; ICP-AES = Inductively Coupled Plasma Atomic Emission Spectroscopy; ICP-MS Inductively Coupled Plasma Mass Spectrometry; IMS = Ion Mobility Spectrometry; IRS = Infrared spectroscopy; LA-ICP-MS = Laser Ablation-Inductively Coupled Plasma Mass Spectrometry; LC = Liquid Chromatography; LC-MS = Liquid Chromatography with Mass Spectrometry; MT = Marquis Test; NFDRs = Non-firearm discharge residues; NMR = Nuclear Magnetic Resonance; PGC = Pyrolysis Mass Chromatography; PLM = Polarized Light Microscopy; PMS = Pyrolysis Mass Spectrometry; PVD = Portable Vapor Detector; RM = Raman Spectroscopy; RR = Resonance Raman Spectroscopy; SEM-EDS = Scanning Electron Microscopy with Energy Dispersive Spectroscopy; SERRS = Surface Enhanced Resonance Raman Scattering Spectroscopy; SIRMS = Stable Isotope Ratio Mass Spectrometry; SFST = Standardized Field Sobriety Tests (SFST); SEM = Scanning Electron Microscopy; ST = Scott test; TLC = Thin Layer Chromatography; VLMS = Visible Light Microspectrophotometry; VOCs = Volatile Organic Compounds; XRD = X-ray powder diffraction; XRF = X-ray Fluorescence.

²**Chemical abbreviations:** HMX = octahydro-1,3,5,7-tetranitro-1,3,5,7-tetrazocine or cyclotetramethylenetetranitramine; MDA = 3,4-methylenedioxyamphetamine; MDMA = 3,4-methylenedioxymethylamphetamine; PETN = pentaerythritol tetranitrate; RDX = cyclotrimethylene-trinitramine; TATB = triamino-2,4,6-trinitrobenzene; tetryl = trinitrophenylmethylnitramine; Δ^9 -THC = Δ^9 -tetrahydrocannabinolic acid; TNT = trinitrotoluene. VOCs = volatile organic compounds.

³**References** of E-nose applications and methods for comparable detections of target compounds (for each forensic sample type) as for conventional methods of analysis. NR = no references found.

Table 1: Identification of components in forensic sample types (human manufactured products and body fluids containing volatile organic or inorganic compounds) using conventional chemical analyses and new potential electronic-nose technologies.

Because certain types of E-nose devices are capable of detecting a wide range of compounds [17,52], they are commonly used to detect hazardous chemicals in the environment including industrial and sanitation wastes (air, water, and soil pollutants) [17], pesticides [62,63], medical wastes [64], and toxins [52]. Forensic sample types containing inorganic materials include firearm discharge residues (FDRs), non-firearm discharge residues (NFDRs), heavy metal toxins, primary explosives, particulates in pigments and extenders, glass, and soil samples [6]. Chemical explosives consist of compounds with inorganic or organic oxidized functional groups such as acetylides, azides, chlorates, fulminates, nitrates, nitrites, ozonides, perchlorates, and peroxides. Thus, E-nose instrument types that detect VICs are potentially useful for gathering forensic evidence when inorganic compounds are involved in criminal fatalities. Most other forensic sample types consist of VOCs from numerous chemical classes.

Conventional analytical methods utilized in forensic sample analyses usually require several different steps and multiple analytical instruments for VOC identifications and for confirmation of sample composition. Each instrument provides a different type of information about the chemical composition and physicochemical characteristics of forensic samples. Consequently, different forensic sample types must be analyzed by different combinations of analytical instruments due to differences in the chemical properties of compounds (analytes) present in the samples and the chemical-detection limitations of analytical instruments as determined by instrument design, operating principles and mechanisms of chemical detections. For example, illegally manufactured drugs must be further analyzed to determine the source or origin of the material, usually based on the occurrence of specific types, concentrations and mixtures of impurities or contaminants within the sample that are unique to a source, batch, or location from which the sample originated. The analysis of the precise types and concentrations of impurities found in forensic samples is referred to as composition profiling or chemical profiling. Chemical profiling of sample impurities is a key method used in the analysis of certain types of explosives, drugs, and trace materials where answering questions concerning origin are important in determining the inv-

olvement of suspects in various criminal activities. Chemical profiling also is necessary for analysis of amphetamines, cocaine, cannabis, and heroin drugs to determine the particular source and batch of these products, manufactured in illegal drug operations. Drugs usually are the most common type of toxic materials analyzed by forensic toxicologists. Besides the analysis of drug impurities, chemical profiling of drugs may also involve the detection of other variations in drug composition such as drug purity, the ratio of actual drug to excipients (tablet bulking, fillers, or dilution agents), degree of hydration, form (acid, base, or salt), as well as the presence of trace alkaloids and isomers [6].

Electronic nose instruments are particularly suited for chemical profiling because these tools provide information about the aroma characteristics of the entire headspace derived from forensic samples including all impurities present. The presence of specific types and combinations of impurities provide important clues about the particular processing methods used to produce the sample such as specific chemical or manufacturing processes, providing an effective means of distinguishing between forensic samples from different sources of origination or manufacture. Thus, chemical profiling-type analyses will likely be among the major roles that E-nose devices will offer and contribute to forensic science and sample analyses in the future. Because many E-nose instrument types are very sensitive to moisture content of the sample, the moisture content of the carrier gas (filtered air) must be controlled and standardized to eliminate variations in E-nose signal output due to water vapor interactions with the sensor array [65].

E-nose detection of human scents

Detections of scents or vapors released from the human body provide very useful information for locating individuals, particularly victims of crimes, and for evaluating the physiological condition (e.g. use and exposure to drugs) and general state of health of individuals involved in crimes, including both victims and perpetrators of crimes. For this reason, E-nose instruments increasingly have been used in the medical industry to facilitate disease diagnoses, obtain assessments of human health in point-of-care patient examinations, and other applications in the biomedical field [3]. Progress in E-nose applications in the medical industry is beginning to spill over into new related applications for forensic science.

One new significant area of potential forensic applications for E-nose instruments is in the detection and location of buried individuals (both living and deceased) as well as the human remains of victims of violent crimes. Individuals who become buried in the rubble of collapsed buildings and other structures as a result of natural disasters (such as earthquakes, floods, avalanches of soil or snow, tornados, hurricanes or other violent natural calamities or weather events) or intentionally buried by violent criminals, generally must be located in a relatively short period of time after the burial event to be successfully rescued from all potential hazardous forces that can threaten a victim's life in such situations. A relatively new approach for detecting living victims that are buried in rubble is to develop chemical detection devices to replace sniffer dogs that usually require frequent rest intervals after periods of active searches. Electronic-nose instruments are not subject to operator fatigue.

Currently, buried human remains most often are detected using ground-penetrating radar (GPR), manual probing techniques, and trained 'cadaver dog' canines. It is not well understood which specific chemicals are detected by cadaver dogs to locate human remains, but the high success rate of trained canines has demonstrated the effective use of human scent as targets of detection. Trained dogs have been very useful in discriminating scents and for detecting explosives, accelerants, narcotics and other drugs, as well criminals and missing persons on foot. Certain canines are capable of discriminating between human remains and other mammals, odors emitted by live individuals, recently deceased, and human remains in various stages of decomposition. Unfortunately, canines used for human-remains detection are a minor portion of the law enforcement canine population due to the high costs associated with the purchase, training, and care of these animals. Nevertheless, canines possess keen olfactory discrimination capabilities that often are far more sensitive and discriminative than many analytical instruments.

Different approaches for detecting and locating living buried victims using E-noses depend on the different types of target compounds intended to be detected based on VOCs released from the victim's bodies in response to stress, oxygen deprivation, excretions, and other adverse conditions associated with being trapped for prolonged periods of time. The bodies of individuals that are subject to variable degrees of suffocation, dehydration, wounding, and starvation or prolonged exposure to adverse elements (temperature extremes, toxic fumes etc.) produce and release different types of gases as a result of various types of metabolic and physiological changes that occur in the body in response to physical afflictions, deprivations and associated stresses. The categories of bodily gases released in association with different adverse conditions are summarized in the upper section of Table 2.

A recent study by Mochalski et al. [93] has revealed some interesting points relating to the sensing of buried human victims. They found that among the VOCs (composing human scent) that serve as potential markers of human presence during Urban Search and Rescue (USR) operations, organized following natural or man-made disasters (e.g. earthquakes, explosions and terrorist attacks), breath volatiles and to a lesser extent skin volatiles are the principal sources of human scent constituents. Their reasoning was that trapped victims have to breathe and that breath constituents, as long-lasting emission sources of VOCs, can help to discriminate between living humans and corpses. They concluded that even though blood and urine in the close vicinity of victims usually offer only temporary sources of human volatiles for detection, these human fluid sources of VOCs should not be underestimated because earthquake and explosion victims frequently are severely injured with blood volatiles comprising a significant important reservoir of human scent VOCs. Consequently, baseline knowledge of all human scent profile constituents, along with the contribution of particular sources in the human scent pool, is critical in order to determine the most appropriate USR sensing targets for E-nose sensing of human bodies trapped by various causes and in different circumstances and conditions.

The recovery of the bodies of victims who die as a result of exposure to adverse conditions or injuries due to burial may be detected with E-noses using a different set of VOCs than are used for buried live victims. The particular types of VOCs released from the decomposing bodies of victims of natural disasters and violent crimes depend on the type of chemical processes involved in decomposition. Some of the major chemical constituents released from human cadavers, produced primarily as a result of microbial decomposition, include such chemicals as cadaverine, putrescine, etc. as indicated in the lower section of Table 2. Very similar chemical classes of VOCs are released from the decomposing bodies and remains (carrion) of large non-human vertebrates, such as dogs and pigs, and the scent markings of wild mammals [94,95].

Applications	E-nose detection	Substrates/ Physiology [†]	Compounds present	Chemicals classes	Example compounds (VICs and VOCs)	References
Location of cadavers and human re- mains	Decomposition (by autolysis)	soft human tissues	478 VOCs (identified by GC-MS)	Oxides Benzene deriv. Aliphatic HC Polycyclic AH Heterocyclic HC Methyl esters Cl-, F- aliphatic HC	Sulfur dioxides dimethyl benzene, toluene undecane naphthalene methenamine hexadecanoic acid methyl ester	[1,2, 66-68]
			Small mol. wt. gases	Oxides Simple HC Sulfides Ammonium	carbon dioxide methane hydrogen sulfide ammonia	[69-72]
	Putrefaction (by microbial action)	Proteins (amino acids)	Aliphatic amines Aromatic, heterocyclic	Diamines Indoles	cadaverine putrescine skatole	[73-77] [78]
		Fatty acids	Short chain organic acids	Carboxylic acids	propionic acid butyric acid	[72,79,80]
Location of live trapped persons	Respiration gases	Inorganics	Small mol. wt. gases	Oxides	carbon dioxide	[81,82]
	Stress compounds	Oxidative stress	Aliphatic HC	Aldehydes	hexanal	[83,84]
		Dehydration				[85]
		Ketosis (starvation)	Aliphatic HC	Ketone	acetone	[86-88]
	Wound com- pounds	Contusions, lacera- tions, ischemia	Aliphatic	Complex VOCs mixture	tritetracontane, nonahexa- contanoic acid, 4-(2,6-di- methyl-1-cyclohexen-1-yl) morpholine	[89,90]
Waste excretion	Urination	Aliphatic	Carbamide	urea	[91,92]	

[†]Number of sensors and sensor type abbreviations: Carbon black composite (CBC), Carbon dioxide sensor (CO₂), Conducting polymer (CP), electrochemical (EC), Metal oxide semiconductor (MOS), Metal oxide semiconductor field effect transistor (MOSFET), Quartz crystal microbalance (QMB), surface acoustic wave (SAW), and Tin dioxide (SnO₂), a type of MOS sensor.

Table 2: Potential E-nose forensic applications for the location of human bodies and remains through the detection of complex gaseous mixtures of VOCs released from these sources under various conditions and situations.

II E-nose detections of breath volatiles

The investigation of chemical indicators (bioindicators) of human metabolism or physiology through a specialized analysis approach, known as metabolic profiling, is a relatively new research area that has received considerable attention due to the potential for simplifying many human-scent related chemical analyses. The investigation of human scents using metabolic profiling is recognized as a way to rapidly and noninvasively detect gaseous mixtures released from the human body that provide significant information about general health, physiological condition, presence of disease, exposure to toxic substances, and many other exogenous factors that influence the outcomes of crime-related events of interest to forensic scientists who are primarily responsible for determining the precise conditions and events that occurred in a criminal case and the factors that affected the ultimate outcome for victims of criminal activities.

Phillips et al. [96] found over 2,000 VOCs in the human breath of healthy individuals using two-dimensional gas chromatography coupled with time-of-flight mass spectrometry (GCxGC-TOF MS), a powerful new tool for multidimensional analysis of complex chemical mixtures. About fifty of these VOCs had the highest alveolar gradients (abundance in breath minus abundance in ambient room air) mostly comprised of benzene derivatives, acetone, methylated alkane derivatives, and isoprene. Some very specific metabolites in the human breath have been highly correlated with certain types of human pathogens, diseases and metabolic disorders [3,97].

Bioindicators of human diseases and causes of death

The specific classes of VOCs comprising the major groups of abnormal chemicals (those not normally found in a healthy body) that are expired in the breath from the body in association with various diseases, genetic disorders, microbial infections (bacterial, fungal, and viral), and metabolic byproducts of microbial degradation of deceased individuals are presented in Table 3. These major groups of abnormal VOCs are released from the human bodies of living patients who are either not in good health or have adverse physiologies as a result of various diseases. Abnormal VOCs often persist in the bodies of postmortem patients for an indefinite period of time following death. Also, the composition of VOCs released from the body changes over time following death, resulting in different metabolic profiles revealed in E-nose analyses. Consequently, the composition and ratio of chemical constituents present in the volatile gases released from corpses over time can serve as useful signatures to help determine the time that has elapsed following death (time since death) or postmortem interval (PMI). The PMI is a useful time reference used by forensic scientists and law enforcement personnel to compare against the activities and whereabouts of potential suspects (and their alibis) and to help identify the victims and perpetrators of criminal activities.

Breath profile analysis

The mixture of chemicals released in the human breath is very complex, contains thousands of VOCs that are constantly changing, and is representative of the large complement of biochemical or physiological processes occurring in the entire body [96]. The composition of expired air in the human breath also varies depending on a person's health status and unique body chemistry. Various metabolic processes within the body produce VOC products that are released into the blood and eventually are passed on to the airways once the blood reaches the lungs. When normal human physiological processes break down or are altered by disease (pathogenesis) or metabolic disorders, the mixture of gases released by the lungs in the breath changes because of the altered chemical pathways resulting from the abnormal metabolic changes caused by these various maladies. Consequently, by frequent monitoring and analyzing the changes in composition and amounts of VOCs present in exhaled breath air, commonly referred to as metabolomics (breathomics) or VOC profiling, it is possible to determine a clinical diagnosis to explain the chemical or biological cause(s) of abnormal alterations in breath-air composition. Boots et al. [219] described some of the currently available methodologies for breath sampling, analysis and data processing with indications of their advantages and potential drawbacks as well as different application possibilities of VOC profiling. They pointed out that until recently, VOC profiling has been applied primarily for diagnostic purposes, but it also may be applied as an analytical or monitoring tool to elucidate the heterogeneity observed in chronic diseases, to study the pathogen(s) responsible for reoccurring infections and to monitor treatment efficacy and progress of healing. Thus, VOC components can serve as individual biomarkers of oxidative stress, inflammation, carcinogenesis and many other diseases. The entire compliment of VOCs in breath also can be chemically analyzed as a whole using electronic noses to produce breath patterns or profiles that can be compared to those of healthy individuals or those with differing physiological histories or exposures to different ambient (atmospheric) environments. Breath profiles produced using E-noses are more useful than those produced from conventional analytical instruments such as GC-MS because E-nose breath profiles can be stored and analyzed as a whole for comparison against application-specific reference (aroma breath profile) databases whereas GC-MS chemical profiles must be analyzed on an individual-compound basis which is considerably more time consuming for routine clinical use where rapid real-time detection and diagnosis is required.

Montuschi et al. [220] found that E-nose breathprints effectively discriminate between patients with different respiratory diseases (including asthma, COPD and lung cancer associated with airway inflammation activity) from healthy control subjects. They also suggested that uses of E-noses could be combined with other '-omics' sensing platform technologies to contribute to the identification of new surrogate markers of pulmonary inflammation and various other respiratory diseases.

Biller et al. [221] evaluated exhaled breath profiles using the Cyranose 320 E-nose, a promising non-invasive diagnostic tool for the discrimination of breath prints between patients with COPD and asthma, to assess whether exhaled breath profile analysis could detect the inflammatory airway response (IAR) induced by ozone inhalation. E-nose signals from exhaled breath profiles showed no significant differences or correlation in the occurrence of IAR between subjects with or without exposure to ozone inhalation. However, independent of ozone exposure, E-nose sensor data did correlate with serum surfactant protein D levels and to a lesser extent with blood neutrophil levels.

One of the biggest technological challenges in developing breath analyzers is to accurately measure a trace amount of VOC analytes in the presence of many interfering gases with a highly concentrated water vapor [222]. Human breath is nearly saturated with water vapor (>95% relative humidity, RH) that often overloads the E-nose sensor array leading to failure of the breath analyzer. This problem can only be solved by proper breath sample conditioning in the mouthpiece before air passes into the sensors for VOC detection. The development of effective and efficient breath-sampling mouthpieces, to filter out interfering water vapor components in the breath, would be very useful in improving VOC breath-analysis methods.

The diagnostic approach of analyzing VOCs in exhaled breath samples constitutes a new frontier in medical diagnostics because it is a noninvasive and potentially inexpensive way to rapidly detect numerous illnesses. Conventional analytical methods for identifying VOCs associated with specific diseases, such as various types of spectroscopy, have shown to be feasible for diagnosing many diseases in all parts of the body using breath-analysis tests, but these traditional approaches require expensive equipment, sophisti-

ated methods, and high levels of expertise to operate effectively in clinical situations. Newer E-nose sensors based on nanomaterials are likely to become the clinical and laboratory diagnostic tools of choice for the future because these instruments are significantly smaller, easier to use, and less expensive than spectrometry or spectroscopy. An ideal nanomaterial-based sensor for breath testing should be sensitive at very low concentrations of VOCs, have a rapid response time, and provide a consistent output for specific mixtures of VOC analytes [223].

The VOCs emitted by the human body have a great potential for medical diagnosis and therapeutic monitoring because their analysis offers a unique insight into biochemical processes ongoing in healthy and diseased humans. Breath analysis holds a distinguished status in this context as it is noninvasive and breath biomarkers can provide valuable information on disease processes, or metabolic disorders occurring even in distant parts of the body away from the lung. Unfortunately, the origin and metabolic fate of numerous breath VOCs have not been elucidated in sufficient depth, thereby limiting the clinical application of breath tests [224]. Consequently, more research is needed to fully elucidate the sources of VOCs in human breath, and establish stronger correlations of bioindicators to specific diseases, so that this information can be used to further advance the application of E-nose technologies to detect specific mixtures of VOCs for disease diagnosis and monitoring.

Disease diagnosis from cell line volatiles

The isolation of diseased human cells in tissue culture lines, as the representative source of tissues where abnormally altered physiological processes are occurring due to disease or metabolic disorders, is another approach for diagnosing diseases using VOCs. In this case, the method is not noninvasive because the removal of tissue samples from the body through surgical biopsies is required to obtain the tissue cell lines for study. This approach is different from breath analysis that usually is noninvasive as a result of simple collection and analysis of VOCs in exhaled breath gases.

Amal et al. [225] built a predictive model to detect metastasis in Hepatocellular carcinoma (HCC), a common and aggressive form of cancer, by using discriminant factor analysis with pattern recognition of VOC fingerprints from HCC cancer and normal cell cultures, analyzed using nanomaterial-based E-nose sensors. The results constitute a proof-of-concept for the in-vitro prediction of the metastatic potential of HCC from VOC fingerprints using nanotechnology that could benefit the development of a fast and potentially inexpensive laboratory test for subclinical HCC metastasis.

Mochalski et al. [224] utilized HepG2 liver cell lines to study VOCs released by the liver which are biomarkers related to products of enzymes involved in drug metabolism (such as cytochrome P450 enzymes). HepG2 is a cell line derived from a 15-year old male patient with a liver carcinoma that possesses an epithelial morphology and secretes a variety of major plasma proteins (e.g., albumin, transferrin and the acute phase proteins fibrinogen, alpha 2-macroglobulin, alpha 1-antitrypsin, transferrin, and plasminogen). The hepatocellular carcinoma (liver cancer) cells were incubated in specially designed headspace 1-L glass bottles sealed for 24 hours prior to headspace analysis. Nine compounds were found to be metabolized and twelve different compounds were released by the carcinoma cells, reflecting the activity of liver enzymes and thus the potential of VOC headspace analysis for the assessment of liver enzyme function.

Pennazza et al. [226] analyzed the VOC mixtures released from well-characterized tumor cells including human melanoma cell lines LOIA, FORM, FIV, a thyroid carcinoma cell line FRO, and synovial sarcoma cell line CME using an E-nose sensor array composed of six crystal quartz microbalance (QMB) sensors, each having a thin surface coating of a different metalloporphyrin to provide differential chemical sensitivity, operating at a 20 MHz resonant frequency. QMB E-noses have quartz crystal resonators that vibrate at a frequency proportional to the mass of the molecules adsorbed to the sensor surfaces. The VOC patterns from the sensor array for the melanoma cell lines were distinctly different from those of the thyroid carcinoma and sarcoma cell lines. Experimental results suggested the possibility of detecting tumors in vivo through the analysis of released VOCs from different body compartments, such as breath and skin.

Single metabolite-specific detection

One of the strong advantages of E-nose detection is the capability of adjusting the chemical sensitivity of individual E-nose sensors in a sensor array in order to tailor the instrument to a very specific narrow range of chemical detection for VOCs in a particular chemical class, or even for a single compound when this is sufficient for detecting certain metabolic or physiological events strongly correlated with release of that compound. Application-specific E-noses with specialized reference databases may focus on a very narrow range of analytes to simplify detection of specific diseases or determine the metabolic health states of organs in the body.

Breath analysis offers the huge potential for early-stage detection and monitoring of diseases to drastically reduce medical diagnostic costs and improve the quality of life of patients suffering from numerous chronic lung illnesses. Righettoni et al. [227] evaluated the detection of the single compound (acetone in the human breath) as a promising noninvasive diagnostic and painless method for monitoring diabetes. A portable E-nose sensor, consisting of flame-deposited and in situ annealed, 10 mol% Si-doped epsilon-WO₃ nanostructured films, was developed with a miniaturized sample chamber volume, sensing temperatures optimized for the low detection limit of acetone (~20 ppb), and short response (10–15 s) and recovery times (35–70 s). Sensor signal (response) was robust in being able to detect and monitor acetone levels continuously at variable exhaled-breath flow rates and at realistic relative humidity ranges (80–90%) in the human breath. This portable experimental nanostructured film sensor device performed comparably to that of state-of-the-art proton transfer reaction mass spectrometry (PTR-MS) and provides an alternative to more elaborate breath analysis techniques. The Si-doped WO₃ nanoparticle sensors were highly selective to acetone over ethanol and had sen-

response times below 15s, making these devices attractive for breath analysis. Acetone concentrations were measured with high signal-to-noise ratios >10.

Rogers et al. [228] demonstrated the use of microsensor-based devices, for detecting select biomarkers in exhaled breath, as a fast and inexpensive breath-screening technology. Micro-hotplate elements with three chemi-resistive metal-oxide films (SnO_2 , In_2O_3 , and CuO) were tested for data acquisition in simulated breath containing single targets [(5 to 20) $\mu\text{mol/mol}$ ammonia, methanol, and acetone], and mixtures of these chemical species. A supervised hierarchical machine-learning algorithm using linear discriminant analysis (LDS) for dimensional reduction of sensing data and discrimination was developed for successful classification and quantification of model biomarkers in validation-set mixtures.

III Emerging E-nose applications

The potential for the development of new biomedical and forensic (cause of death) applications using E-nose instruments is high given the rapid progress in correlating VOC bioindicators and breath gas profiles to specific causes of disease, death, and health conditions of crime victims. New E-nose devices are being developed with the capability of detecting specific types and patterns of VOC profiles for numerous applications. Barash et al. [229] recently developed a gold nanoparticle (GNP) gas sensor E-nose that could distinguish between healthy lung cells and diseased cells with small-cell lung cancer (LC), non-small cell LC, adenocarcinoma or squamous cell carcinoma. This instrument has the potential to revolutionize LC screening as well as early and differential diagnosis of LC subtypes of unreachable lung nodules based on specific patterns of VOC profiles derived from the analysis of headspace from LC cells. In a similar study, Broza et al. [230] demonstrated the feasibility of using a nanomaterial-based E-nose sensor to identify the breath-print of early stage LC and to assess the difference in LC states before and after lung surgery to remove tumor tissue. They found five VOCs that were significantly reduced after LC surgery (lung resection). Tisch et al. [231] noted that most lung cancers originate from epithelial cells that undergo genetic mutations leading to changes in protein levels and post-translational protein modifications that presumably generate changes in VOCs (relative to healthy, unmutated epithelial cells) that are released in exhaled air. Xu et al. [232] examined the feasibility of using a non-invasive nanomaterial-based breath test to replace upper digestive endoscopy and biopsy for the detection of gastric cancer.

Several studies have provided evidence to show the potential for using breath biomarkers to detect and diagnose active bacterial infections in the lung. Phillips et al. [211] evaluated breath VOC biomarkers in subjects with active pulmonary tuberculosis (TB), caused by *Mycobacterium tuberculosis*. They found that metabolic products of *M. tuberculosis*, principally derivatives of naphthalene, benzene, and alkanes, could be reliably used to detect this pathogenic bacterium in subjects with active TB using a six-minute point-of-care breath test developed to detect these TB-specific volatile biomarker metabolites. Španěl and Smith [233] found that hydrogen cyanide was released by *Pseudomonas* bacteria into the breath of children with cystic fibrosis. They also found other correlations for biomarkers such as the presence of breath acetone that varied with diet, ammonia as an indicator of dialysis efficiency, and hydrogen and CO_2 levels that were related to gastric emptying and bowel transit times. Michael et al. [234] suggested that future bedside VOC profiling would probably enable the rapid characterization of microbe-associated diseases to facilitate diagnosis and treatments by healthcare practitioners. They observed that VOCs are indicative of both healthy and disease states because VOC profiles, for any given anatomical site in the body, are dependent on VOCs produced by both human tissue (host component) and any microbes that are present in these same tissues.

The results from many studies have shown the capability of various E-noses to distinguish between different types of lung diseases. Fens et al. [158] showed that a new experimental E-nose could be used to discriminate between asthma and fixed airways COPD via differences in exhaled breath profiles. Many other examples, demonstrating the use of E-noses to distinguish between different diseases, are listed among the references in Table 3.

The repeated E-nose monitoring of breath gas profiles from individuals has been shown to indicate changes in body biochemistry and health state, providing a means of determining if a person is recovering or getting worse due to particular ailments. Fuchs et al. [235] found that isoprene (2-methylbuta-1,3-diene) represents a precursor to isoprenoid and cholesterol biosynthesis and that a decline in exhaled isoprene in LC patients was correlated with immune activation which they surmised was related to changes in lipid metabolism.

Numerous studies have provided a large amount of evidence to show that the composition of exhaled breath is affected by a person's exposure to exogenous chemicals in inhaled air such as through smoking habits, living near the source of air pollutants, or exposure to smoke from fires. Španěl et al. [236] found that all inhaled exogenous compounds are partially retained in the exhaled breath. Through this understanding, the biochemical background or history of inhaled chemical exposures (from various recent prior exposure events) can be deduced to determine the current state of health of victims, cause of death (through inhalations of toxic gases), or exposure to air pollution, smoke from fires, or indoor-air contaminants. This information provides valuable indications of the prior location of victims, (relative to crime scenes) based on exposure to local or point-sources of pollutants or toxic substances. This information is useful for determining the effects of inhalation factors on crime incidences and whether a victim was moved from the crime scene. Filipiak et al. [237] found that the composition of exhaled breath is considerably influenced by exposure to airborne pollutants, contaminants, and particularly by smoking. They found 80 VOCs that were significantly related to smoking, and suggested that the proper interpretation and full understanding of breath profile data required a careful investigation of the potential biological and chemical origins of breath volatiles, either from endogenous or exogenous sources.

Disease/Disorder/ Injury/Infection ¹	Organ	Biomarker indicator VOCs ²	References
AFDL	Liver	Acetaldehyde, isoprene, other VOCs	[98]
AHI	Liver	Ethane, pentane (volatile alkanes)	[99]
ALF	Liver	Complex VOCs profile	[100]
ARDS	Lung	Acetone, isoprene, n-pentane	[101,102]
Aspergillosis (invasive)	Lung	2-pentylfuran	[103]
		Complex VOCs profile	[104]
Asthma	Lung	Pentane, ethane, isoprene	[105-107]
		Leukotriene B4, prostaglandin E2	[108,109]
		8-isoprostane	[110]
		Nitric oxide	[111]
		Complex VOCs profile	[112-116]
BCKD	Systemic, Kidney	2-oxoisocaproic acid	[73]
Cancer	Bladder	Complex VOCs profile	[117]
	Breast	C4-C20 alkanes, monomethylated alkanes	[118]
	Head and neck	4,6-dimethyl-dodecane, 2,2-dimethyl-propanoic acid, 5-methyl-3-hexanone, 2,2-dimethyl-decane, limonene, 2,2,3-erimethyl-, exobicyclo[2.2.1]heptane	[119]
		Dimethyl trisulfide	[120]
	Lung	Alkanes, monomethylated alkanes	[121,122]
		Alkanes, aromatic compounds	[123]
		Aniline, o-toluidine	[124]
		Aliphatic aldehydes	[125,126]
		1-Butanol, 3-Hydroxy-2-butanone	[127]
		Dimethyl sulfide, dimethyl formamide, butane, butanal	[128]
		Ethane	[129]
		Isoprene, acetone, methanol	[130]
		Pentane	[131]
		1-octene	[132]
	Complex VOCs profile	[73,121,130, 131,133-148]	
Skin	Isoamyl alcohol, dimethyldisulfide, trisulfide	[149]	
Chronic hepatitis	Liver	Methyl-mercaptan, dimethyl sulfide	[150]
CIP	Lung	Acetone, isoprene, n-pentane	[101]
COPD	Lung	Aldehydes, nitrotyrosine, cytokines	[151]
		Leukotriene B4, 8-isoprostane	[152]
		Hydrogen peroxide	[153,154]
		Nitrate	[155]
		Nitric oxide	[156]
		Ethane	[157]
		Complex VOCs profile	[116,158-164]
CPD	Heart, Lung	Ethanol, acetone	[165]
Cystic fibrosis	Lung	Leukotriene B4, interleukin-6	[166]
		Nitrotyrosine	[151]

		Pentane, propane, methanol, ethanol, acetone, benzene	[167]
		Carbonyl sulphide, dimethyl sulphide, carbon disulfide	[167,168]
		Methyl thiocyanate	[169]
		Ethane	[167,170]
		Isoprene	[167,171]
		Hydrogen cyanide	[172,173]
		Complex VOCs profile	[167,174,175]
Cystinuria	Kidney, ureter, bladder	Cadaverine, piperidine, putrescine, pyrrolidine	[73,74]
Diabetes mellitus	Systemic	Acetone, ethanol, methyl nitrate, complex VOCs	[176,177]
Emphysema	Lung	Complex VOCs profile	[178]
Endocarditis (infective)	Heart	Hydrogen sulfide, methyl mercaptan, dimethyl sulfide	[179-182]
Foetor hepaticus	Liver	Complex VOCs profile	[183]
GERD	Esophagus	Complex VOCs profile	[162]
Hepatic cirrhosis	Liver	Dimethyl sulfide, hydrogen sulfide, mercaptans, fatty acids	[184]
		Dimethyl sulfide, acetone, 2-pentanone and 2-butanone	[185]
		Methyl-mercaptan, dimethyl sulfide	[150,186]
Hepatic coma	Liver	Methyl-mercaptan, dimethyl sulfide	[150,186,187]
Histidinemia	Systemic	2-imidazolepyruvic acid, 2-imidazolelactic acid, 2-imidazoleacetic acid	[73]
Hyperglycemia	Systemic	Methyl nitrate, xylene, ethylbenzene	[177,188]
IBD	Intestine	Pentane, ethane, propane	[189-192]
IHD, angina	Heart	Alkanes, methylated alkanes	[122,193]
ILD	Lung	Ethane	[194,195]
Ketosis, starvation	Systemic	Acetone	[86]
MPM	Lung	Complex VOCs profile	[196-198]
NSCLC	Lung	1-Butanol, 3-Hydroxy-2-butanone	[127]
		Complex VOCs profile	[142,147]
Oxidative stress	Systemic	8-Isoprostane	[110]
		Alkanes, methylated alkanes	[199]
PCD	Respiratory tract	Complex VOCs profile	[175]
Phenylketonuria	Systemic	Phenylpyruvic acid, phenyllactic acid, phenylacetic acid	[73]
PLC	Lung	Formaldehyde, propanol, isoprene, acetone, o-toluidine	[200]
Renal dysfunction	Kidney	Complex VOCs profile	[201]
Respiratory infections	Lung	Complex VOCs profile	[202]
Rheumatoid arthritis	Bone joints, cartilage	Pentane	[203]
Schizophrenia	Brain	Pentane, carbon disulfide, ethane	[204-206]
SFS	Feet	Butyric acid, hexanoic acid; trans-3-methyl-2 hexenoic acid	[207]
TB	Lung	Methyl nicotinate	[208]
		Complex VOCs profile	[86,209-213]
Tyrosinemia	Systemic	p-hydroxyphenylpyruvic acid	[73]
Upper respiratory infections	Respiratory tract	Acetic acid, acetaldehyde, 2-butene, methyl methacrylate, 2,3-butanedione, 2-butenal, vinyl butyrate	[214]
VAP	Lung	Complex VOCs profile	[215-218]

¹**Abbreviations:** AFDL = Alcoholic Fatty Liver Disease; AHI = Alcohol-induced Hepatic Injury; ALF = Acute Liver Failure; ARDS = Acute Respiratory Stress Syndrome; BCKD = Branched-Chain Ketoaciduria Disorder (Maple Syrup Urine Disease); CIP = Critically Ill Patients; CPD = Cardiopulmonary Disease; COPD = Chronic Obstructive Pulmonary Disease; GERD = Gastro-esophageal reflux disease; IBD = Inflammatory Bowel Disease; IHD = Isochemic Heart Disease; ILD = Interstitial Lung Disease (includes cryptogenic organizing pneumonia, idiopathic pulmonary fibrosis, sarcoidosis, etc.); MPM = Malignant Pleural Mesothelioma; PCD = Primary Ciliary Dyskinesia; PLC = Primary Lung Cancer; NSCLC = Non-Small Cell Lung Cancer; SFS = Sweaty Feet Syndrome; TB = pulmonary Tuberculosis; VAP = Ventilator Associated Pneumonia.

²Bioindicators are primarily volatile organic compounds (VOCs), initially identified using gas chromatography-mass spectroscopy (GC-MS), or similar analytical instruments, prior to development of corresponding E-nose reference libraries and VOC profiles for application-specific detections.

Table 3: Potential electronic-nose diagnoses of human organ-related diseases and postmortem causes of death through the detection of volatile biomarker indicator compounds in the human breath, exhaled breath condensate, bronchi, or alveolar air.

Recent advancements in methods, used to help improve E-nose analyses (correlating breath bioindicators to E-nose patterns) through identification of breath gas VOC components and sampling methods and models, are important in the development of future E-nose applications to forensic science. Filipiak et al. [238] developed an automated adsorption needle trap method to pre-concentrate breath VOCs from critically ill patients in intensive care to improve sensitivity and reproducibility of NT-GC-MS analysis, also applicable to E-nose analyses. Gilchrist et al. [173] investigated the use of three different bag materials, (Nalophan of 25 µm and 70 µm thickness, and Tedlar), for collection and storage of breath-derived samples containing hydrogen cyanide. The latter two bag types performed best, retaining HCN concentrations for up to 24 h. Mochalski et al. [239] investigated the stability of 41 selected VOC breath constituents in three types of polymer sampling bags and found that Tedlar bags were superior to Kynar and Flexfilm sampling bags in terms of background emission, chemical species stability, and reusability.

Ibrahim et al. [115] generated a breath analysis model based on 15 VOCs to classify asthma patients with an accuracy of 85%. This non-invasive disease phenotyping model could lead to clinical application for classifying asthma patients into sputum, neutrophila, and uncontrolled asthma phenotypes. King et al. [240] produced a similar model for the evaluation of isothermal rebreathing, an experimental technique for estimating the alveolar air levels of hydrophilic VOCs in exhaled breath gases. This model clarifies the discrepancy between in vitro and in vivo blood-breath ratios of hydrophilic VOCs and helped to explain the exhalation kinetics of exchange between blood-borne and exhaled breath VOCs. King et al. [241] previously had developed a mathematical method for the sampling of other trace gases in exhaled breath, especially VOCs like acetone that reflect ongoing metabolism. Koc et al. [242] developed the first mathematical model for isoprene in exhaled breath that provides supportive evidence for a peripheral (extrahepatic) source of isoprene, the most abundant exogenous VOC contained in human breath which is considered a potentially useful biomarker for diagnostic and monitoring purposes. Martinez-Lozano [243] utilized secondary electrospray ionization mass spectrometry (EIMS) to quantify and identify the abundance of carboxylic acids (organic acids) in the breath following sucrose intake. Rapid increases in the concentrations of propionic and butanoic acids in the breath were attributed to bacterial activity in the mouth and pharynx. Carboxylic acids in the breath are readily detectable by certain E-nose instruments and could be used to diagnose bacterial infections in the lung, upper respiratory tract, as well as in the mouth and throat.

Phillips et al. [161] examined machine-learning approaches to analyze breath data for the diagnosis of COPD in patients based on unique combinations of VOCs found in the breath. They found that a patient's smoking status affected COPD-classification, requiring cross-validation with appropriate controls. Ulanowska [144] applied statistical methods, such as discriminant analysis (DA) and the CHAID model tree, to breath-profile data in order to identify patients with lung cancer. Their results indicated that patients with lung cancer had higher concentrations of certain VOCs (ethanol, acetone, butane, dimethyl sulfide, isoprene, propanal, 1-propanol, 2-pentanone, furan, o-xylene, and ethyl benzene) compared to healthy nonsmokers. A few other VOCs (pentanal, hexanal, and nonane) were found only in the breath of people who suffered from cancer. They also discovered higher concentrations of acetonitrile, benzene, and furan derivatives in nonsmokers. DA showed that butyrolactone, carbon disulfide, and dimethyl sulfide had to be considered in breath analysis in order to definitively recognize and distinguish between healthy subjects (with different smoking habits) from those suffering from cancer.

Filipiak et al. [218] identified specific pathogen-derived volatile biomarkers in breath that could be used for the early and noninvasive diagnosis of ventilator associated pneumonia (VAP). In vitro experiments using cultures of bacteria most frequently associated with VAP patients, i.e. *Staphylococcus aureus* and *Pseudomonas aeruginosa*, were performed to investigate the release or consumption of specific VOCs associated with these species. They found many distinct differences in VOCs released from cultures of these two bacteria for aldehydes, carboxylic acids, alcohols, ketones, hydrocarbons, esters, volatile sulfur compounds (VSCs) and volatile nitrogen compounds (VNCs) chemical classes. The results provided strong evidence to suggest that the detection and identification of pathogenic bacteria could be achieved by determination of characteristic volatile metabolites useful in clinical breath-gas analysis as a non-invasive method for the early detection of bacterial lung infections.

IV Confirming E-nose analyses

Electronic-nose instruments could potentially be used in combination with other forensic instruments in several ways. E-nose devices could be used for the initial testing of forensic samples to provide a preliminary indication of the chemical nature of the VOCs present. This information can be used to help direct the types of subsequent analytical tests that need to be performed using conventional analytical instruments. New multiple-detector E-nose instruments are being developed with the capability of simultaneous detection of multiple types of volatile gases [5]. Other instruments with E-nose detectors interfaced in tandem with analytical instruments, similar to GC-MS and LC-MS, are possible. E-noses also may be used to confirm diagnoses and interpretations of chemical analyses made from determinations using conventional analytical instruments [3,4].

V Admission of E-nose evidence in criminal litigations

Forensic evidence based on analytical data from E-nose devices hitherto has not been introduced, used, or admitted into the evidentiary record at any significant level for criminal litigations in the United States. There are also many other analytical methods, still considered in the experimental or unproven stage, that have not yet been established as proven and completely reliable to the extent that they are recognized as acceptable evidence for routine admittance in US courts; such as is currently recognized for numerous molecular biology methods that provide many types of DNA evidence.

In 2000, Rule 702 of the Federal Rules of Evidence for assessing the admissibility of scientific expert testimony was amended to include the Daubert standard. The Daubert standard provides a rule of evidence regarding the admissibility of expert witnesses' testimony (based on analytical methods) in United States federal legal proceedings. A Daubert motion, usually introduced by a defense lawyer, is a special case of motion raised before or during a trial to exclude the presentation of unqualified evidence to the jury. Once certain types of scientific evidence (derived from specific methods or instruments) have been excluded by a Daubert motion because they fail to meet relevancy and reliability standards, they can be challenged when introduced again in another trial as testimony or evidence based on the method. Even though a Daubert motion is not binding in other courts of law, other judges may choose to follow that precedent if certain types of scientific evidence are found untrustworthy by a different court. The U.S. Supreme Court listed some guidelines to help in evaluating the soundness of novel science methods used in forensic analyses. The Supreme Court agreed that before scientific evidence could be admitted as scientific expert testimony, the following principles should apply: 1) the trial judge is the gatekeeper who must assure that scientific expert testimony truly proceeds from scientific knowledge; 2) the trial judge must ensure that the expert's testimony is relevant and rests on a reliable foundation, and the expert testimony cannot be simply referred to the jury as a question of weight; 3) a conclusion will qualify as scientific knowledge if the proponent can demonstrate that it is a product of sound scientific methodology derived from the scientific method; in which 4) the scientific methodology is defined as: a) the process of formulating hypotheses and then conducting experiments to prove or falsify the hypothesis, b) that the hypothesis is considered relevant for establishing the "validity" of scientific testimony based on empirical testing (whether the theory or technique is falsifiable, refutable, and/or testable), and c) the method has been subjected to peer review and publication, has a known or potential error rate, has maintenance standards and controls concerning its operation, and the theoretical basis of the technique is generally accepted by a relevant scientific community.

The Supreme Court further ruled that nothing in the Federal Rules of Evidence governing expert evidence "gives any indication that 'general acceptance' is a necessary precondition to the admissibility of scientific evidence". By requiring experts to provide relevant opinions grounded in reliable methodology, proponents of Daubert were satisfied that these standards would result in a fair and rational resolution of the scientific and technological issues. The Supreme Court explicitly cautioned that the Daubert list should not be regarded by judges as "a definitive checklist or test...". Yet in practice, judges have judged the admissibility of scientific evidence using the "Daubert factors" as a checklist. Even though the Daubert standard is now the law in federal courts and in over half of U.S. states, the Frye standard remains the law in some jurisdictions including California, Illinois, Maryland, New York, New Jersey, Pennsylvania, and Washington state.

The absence of agreed upon protocols for the validation of scientific techniques, prior to their being admitted in court, has been considered entirely unsatisfactory and unfair in courts of law. Judges are not well qualified to determine scientific validity without input from scientists. Thus, some countries have recommended that a Forensic Science Advisory Council be established to develop "gate-keeping" tests for expert evidence. This process should be accomplished in partnerships between judges, scientists and other key players in the criminal justice system. In 2005, the United Kingdom House of Commons Science and Technology Committee recommended the creation of a Forensic Science Advisory Council to regulate forensic evidence in the UK. A similar science-based advisory council would be very useful and instrumental in establishing the validity of data and evidence from new, emerging scientific methods and technologies in the United States. The Law Commission for England and Wales has proposed a consultation paper (No.190) to adopt a criterion like the Daubert Standard to help reform the law of evidence in regards to the admissibility of scientific evidence.

Thus, the validation of E-nose evidence, based on standardized methods of acquisition, must first be established with reliable and consistent standardized methods and with Daubert-standard certifications to assure the strength and validity of E-nose data in criminal investigations and court proceedings. Once E-nose methods have been validated and introduced as reliable evidence with increasing frequency in criminal litigations, these instruments should provide valuable additions to the tools available to forensic scientists of the future.

VI Future potential E-nose applications

Human breath analysis, a promising new field of medicine and medical instrumentation, potentially offers noninvasive, real-time, point-of-care (POC) disease diagnostics and metabolic status monitoring for many illnesses [244]. Numerous breath biomarkers were detected and quantified previously using GC-MS techniques [245], Proton Transfer Reaction MS (PTRMS) [246], and selected ion flow tube mass spectrometry, SIFT-MS [247]. Recently, high-sensitivity laser spectroscopic techniques, including tunable diode laser absorption spectroscopy (TDLAS), cavity ringdown spectroscopy (CRDS), integrated cavity output spectroscopy (ICOS), cavity enhanced absorption spectroscopy (CEAS), cavity leak-out spectroscopy (CALOS), photoacoustic spectroscopy (PAS), quartz-enhanced photoacoustic spectroscopy (QEPAS), and optical frequency comb cavity-enhanced absorption spectroscopy (OFC-CEAS) have been reported [244].

Santonico et al. [248] simultaneously tested the validity of two different E-nose instruments on selected target compounds. A gas sensor array based on the quartz crystal microbalance E-nose (ROTV E-nose) with transducers functionalized with metalloporphyrins, and a Cyranose E-nose were used simultaneously in calibration tests to demonstrate that limits of detection down to tens of ppb are possible. This study provided the first steps towards quality assurance of E-nose data for use in the biomedical field.

Valera et al. [249] evaluated the potential application of a new E-nose for the diagnosis of respiratory tract diseases. It is a simple, portable instrument that may be easily used in daily practice. Other positives include quick results and high reproducibility between instruments over different days. For definitive implementation of this new tool, additional studies are necessary with sufficiently large case volumes in order to determine the more specific VOC patterns of each disease.

The diagnostic accuracy of a sophisticated experimental E-nose (DiagNose, C-it BV) using exhaled air to detect tuberculosis was recently tested [212]. The DiagNose uses a measurement method that enables transfer of calibration models between devices thus eliminating the most common pitfall for large scale implementation of E-noses in general. The portability and fast time-to-result of the DiagNose provides a proactive screening search for new TB cases in rural areas, without the need for highly-skilled operators or a hospital center infrastructure.

Other future potential new E-nose applications include detection of head trauma severity associated with athletic physical-contact injuries, heart disease (such as infective endocarditis) based on the presence of resident oral bacterial populations (releasing halitosis-related VOCs), and other postmortem analyses for specific information relating to causes of death (disease, physiological or genetic disorder, toxins, poisons, physical trauma, drug related, organ failure, injuries) or time of death.

Conclusions

E-nose devices are relatively new analytical tools that may soon be added to the arsenal of methods and techniques useful to forensic scientists and investigators in recreating crime scenes and events based on chemical evidence derived from the analysis of many different types of crime-scene samples that release volatile gases. E-nose instruments potentially offer new types of evidence and provide additional information that can be used in combination with data and evidence collected from conventional analytical instruments utilized in the chemical analysis of forensic samples.

Even though exhaled breath analysis is the least invasive diagnostic method, it is still not yet the preferred, routine method used in clinical practice. The gap standing between breath printing and disease diagnosis is mainly due to the complexity of the many variables affecting the composition of breath gases and the huge variety of available techniques that are still largely confined to research. Bridging this gap will require standardization of sample collection methods, sensor technology and data analysis. Narrowing the gap will require cooperation between researchers and healthcare professions to agree on a unified path for breath analysis methodologies through development of common technological platforms and a shared list of standard operating procedures [250].

The pattern of exhaled breath VOCs represents a person's whole-body metabolic signature (overall physiological health condition) with the potential for identifying and characterizing different types of human diseases including lung cancers. A breath biosignature-based classification of homogeneous subgroups (types) of lung cancer may be more accurate than a global breath signature [148]. Thus, more application-specific E-nose referenced databases of breath profiles used for the single application intended, such as for a specific type of cancer, will usually provide more effective and reliable diagnoses, better predictive results, greater reproducibility (precision), and significant reductions in false positive determinations. Application-specific reference databases (breath profiles) for specific diseases are constructed from the analysis of breaths from subjects with confirmed known diagnoses for each corresponding type of disease (such as lung cancers) included in the reference database. Broader-based reference breath-profile libraries could be constructed for various lung diseases or other diseases of the body. Broader reference databases are useful for initial diagnoses to narrow down the list of possible causes to explain current symptoms and to provide a strategy for subsequent diagnostic tests.

This review has described some potential new methods and solutions that are needed to improve and standardize the processes used in the analysis of breath profiles and VOCs, including the greater integration and utilization of E-nose devices. Applications of E-nose instruments will no doubt lead to even more effective early detections of human diseases and metabolic disorders as scientific breakthroughs in knowledge of bioindicator compounds, correlations with disease incidence, and improvements in gas-detection methods advance with new research. All of these biomedical applications are potentially useful in forensic science investigations to help solve crimes requiring information relating to human health and causes of death.

Acknowledgments

The author appreciates Dr. Viorica Lopez-Avila, research scientist with Agilent Technologies, for the opportunity and invitation to write this review article on the relatively recent developments of potential E-nose applications for the EAD analysis of forensic samples and VOC profiles in the human breath. The author also thanks Mrs. Charisse Oberle for the collection of references and proofreading of manuscript drafts.

References

1. Barshick SA, Griest WH, Vass AA (1997) Electronic aroma technology for forensic and law enforcement applications. *SPIE* 2941: 63-74.
2. Vass AA, Barshick SA, Sega G, Caton J, Skeen JT, et al. (2002) Decomposition chemistry of human remains: A new methodology for determining the postmortem interval. *J Forensic Sci* 47: 542-53.
3. Wilson AD, Baietto M (2011) Advances in electronic-nose technologies developed for biomedical applications. *Sensors* 11: 1105-76.
4. Wilson AD (2011) Future applications of electronic-nose technologies in healthcare and biomedicine. Chapter 15 in *Wide Spectra of Quality Control*, InTech Pub, Rijeka, Croatia.

5. Wilson AD, Baietto M (2009) Applications and advances in electronic-nose technologies. *Sensors* 9: 5099-148.
6. Rendle DF (2005) Advances in chemistry applied to forensic science. *Chem Soc Rev* 34: 1021-30.
7. Jasper JP, Edwards JS, Ford LC, Corry RA (2002) Putting the arsonist at the scene: "DNA" for the fire investigator? *Gas chromatography/isotope ratio mass spectrometry. Fire Arson Investig* 51: 30-4.
8. Brudzewski K, Osowski S, Markiewicz T, Ulaczyk J (2006) Classification of gasoline with supplement of bio-products by means of an electronic nose and SVM neural network. *Sens Actuat B* 113: 135-41.
9. Conner L, Chin S, Furton KG (2006) Evaluation of field sampling techniques including electronic noses and a dynamic headspace sampler for use in fire investigations. *Sens Actuat B* 116: 121-29.
10. Sobanski T, Szczurek A, Nitsch K, Licznerski BW, Radwan W (2006) Electronic nose applied to automotive fuel qualification. *Sens Actuat B* 116: 207-12.
11. Wiziack NKL, Catini A, Santonico M, D'Amico A, Paolesse R, et al. (2009) A sensor array based on mass and capacitance transducers for the detection of adulterated gasolines. *Sens Actuat B* 140: 508-13.
12. Robertson J, Grieve M (1999) *Forensic examination of fibres*. Taylor & Francis Forensic Science Series, London & New York.
13. White PC (2000) SERRS spectroscopy - A new technique for forensic science? *Sci Justice* 40: 113-19.
14. Huang M, Yinon J, Sigman ME (2004) Forensic identification of dyes extracted from textile fibers by liquid chromatography mass spectrometry (LC-MS). *J Forensic Sci* 49: 238-49.
15. Hobbs AL, Admirall JR (2003) Trace elemental analysis of automotive paints by laser ablation inductively coupled plasma mass spectrometry (LA-ICP-MS). *J Anal Bioanal Chem* 376: 1265-71.
16. Schiffman SS, Gutierrez-Osuna R, Nagle HT (2002) Measuring odor intensity with e-noses and other sensor types. *Proceedings of the 9th International Symposium on Olfaction and Electronic Nose, Rome, Italy*.
17. Wilson AD (2012) Review of electronic-nose technologies and algorithms to detect hazardous chemicals in the environment. *Proc Technol* 1: 453-63.
18. Saferstein R (2004) *Criminalistics: An Introduction to Forensic Science*, 8th Edition, Pearson Prentice Hall, New Jersey, USA.
19. Anonymous (2001) *Forensic Examination of Glass and Paint: Analysis and Interpretation*. Taylor & Francis Forensic Science Series, New York, USA.
20. Pye K, Croft DJ (2004) *Forensic Geosciences: Principles, Techniques & Applications*, Geological Society Special Publication, London, England.
21. De Cesare F, Di Mattia E, Pantalei S, Zampetti E, Vinciguerra V, et al. (2011) Use of electronic nose technology to measure soil microbial activity through biogenic volatile organic compounds and gases release. *Soil Biol Biochem* 43: 2094-107.
22. Carrasco A, Saby C, Bernadet P (1998) Discrimination of Yves Saint Laurent perfumes by an electronic nose. *Flavour Fragr J* 13: 335-48.
23. Branca A, Simonian P, Ferrante M, Novas E, Negri RM (2003) Electronic nose based discrimination of a perfumery compound in a fragrance. *Sens Actuat B: Chem* 92: 222-27.
24. Toal SJ, Trogler WC (2006) Polymer sensors for nitroaromatic explosives detection. *J Mater Chem* 16: 2871-83.
25. Boeck GD, Wood M, Samyn N (2002) Recent applications of LC-MS in forensic science. *Recent Appl LC-MS Nov*: 1-8.
26. Wagner E, Clement S (2001) Surface enhanced resonance Raman scattering (SERRS) spectroscopy - study on inks. *Z Zagadnien Nauk Sadowych* 46: 437-41.
27. Laporte GM, Wilson JD, Cantu AA, Mancke SA, Fortunato SL (2004) The identification of 2-phenoxyethanol in ballpoint inks using gas chromatography/mass spectrometry - relevance to ink dating. *J Forensic Sci* 49: 155-9.
28. Locicero S, Dujourdy L, Mazzella W, Margot P, Lock E (2004) Dynamic of the ageing of ballpoint pen inks: quantification of phenoxyethanol by GC-MS. *Sci Justice* 44: 165-71.
29. Van Deventer D, Mallikarjunan P (2002) Optimizing an electronic nose for analysis of volatiles from printing inks on assorted plastic films. *Innov Food Sci Emerg Technol* 3: 93-9.
30. Mensing J, Wisitsoraat A, Tuantranont A, Kerdcharoen T (2013) Inkjet-printed sol-gel films containing metal phthalocyanines/ porphyrins for opto-electronic nose applications. *Sens Actuat B* 176: 428-36.
31. Jenkins AJ (2001) Drug contamination of US paper currency. *Forensic Sci Int* 121: 189-93.
32. Deshmukh S, Jana A, Bhattacharyya N, Bandyopadhyay R, Pandey RA (2014) Quantitative determination of pulp and paper industry emissions and associated odor intensity in methyl mercaptan equivalent using electronic nose. *Atmos Environ* 82: 401-9.
33. Yinon J (2003) *Advances in forensic applications of mass spectrometry*. CRC Press, Boca Raton, USA.
34. Eiceman GA, Stone JA (2004) Ion Mobility Spectrometers in National Defense. New uses of previously unheralded analytical instruments. *Anal Chem* 76: 390A-97A.
35. Evans CS, Sleeman R, Luke J, Keely BJ (2002) A rapid and efficient mass spectrometric method for the analysis of explosives. *Rapid Commun Mass Spectrom* 16: 1883-91.
36. Cullum HE, McGavigan C, Uttley CZ, Stroud MAM, Warren DC (2004) A second survey of high explosive traces in public places *J. Forensic Sci* 49: 684-90.
37. Capua E, Cao R, Sukenik CN, Naaman R (2009) Detection of triacetone triperoxide (TATP) with an array of sensors based on non-specific interactions. *Sens Actuat B* 140: 122-127.
38. Sekhar PK, Brosha EL, Mukundan R, Linker KL, Brusseau C, et al. (2011) Trace detection and discrimination of explosives using electrochemical potentiometric gas sensors. *J Hazard Mater* 190: 125-32.
39. Brudzewskia K, Osowska S, Pawlowski W (2012) Metal oxide sensor arrays for detection of explosives at sub-parts-per million concentration levels by the differential electronic nose. *Sens Actuat B* 161: 528-33.
40. Amania M, Chu Y, Watermana KL, Hurley CM, Platek MJ, et al. (2012) Detection of triacetone triperoxide (TATP) using a thermodynamic based gas sensor. *Sens Actuat B* 162: 7-13.
41. Patil SJ, Duragkar N, Rao VR (2014) An ultra-sensitive piezoresistive polymer nano-compositemicrocantilever sensor electronic nose platform for explosive vapor detection. *Sens Actuat B* 192: 444-51.

42. Brian DP, Seung JL, Martin M, Meinhart CD (2012) Free-surface microfluidics/surface-enhanced Raman spectroscopy for real-time trace vapor detection of explosives. *Anal Chem* 84: 9700–5.
43. Young RC, Buttner WJ, Linnell BR, Ramesham R (2003) Electronic nose for space program applications. *Sens Actuat B Chem* 93: 7–16.
44. Tung TT, Castro M, Feller J-F, Kim TY, Suh KS (2013) Hybrid film of chemically modified graphene and vapor-phase-polymerized PEDOT for electronic nose applications. *Organic Electron* 14: 2789–94.
45. Zeichner A (2003) Recent developments in methods of chemical analysis in investigations of firearm-related events. *Anal Bioanal Chem* 376: 1178–91.
46. Cavicchi RE, Walton RM, Aquino-Class M, Allen JD, Panchapakesan B (2001) Spin-on nanoparticle tin oxide for microhotplate gas sensors. *Sens Actuat B* 77: 145–54.
47. Barnes BB, Snow NH (2012) Recent advances in sample preparation for explosives reference module in chemistry, molecular sciences and chemical engineering comprehensive sampling and sample preparation, analytical techniques for scientists, Vol. 3: Extraction techniques and applications: biological/medical and environmental/forensics. Academic Press, Waltham, Massachusetts.
48. Ceto X, O'Mahony AM, Wang J, del Valle M (2013) Simultaneous identification and quantification of nitro-containing explosives by advanced chemometric data treatment of cyclic voltammetry at screen-printed electrodes. *Talanta* 107: 270–6.
49. Trombka JJ, Schweitzer J, Selavka C, Dale M, Gahn N, et al. (2002) Crime scene investigations using portable, non-destructive space exploration technology. *Forensic Sci Int* 129: 1–9.
50. Sironi S, Capelli, L, Céntola P, Del Rosso R, Grande MI (2007) Continuous monitoring of odours from a composting plant using electronic noses. *Waste Manag* 27: 389–97.
51. Santra S, Guha PK, Ali SZ, Hiralal P, Covington JA, et al. (2010) ZnO nanowires grown on SOI CMOS substrate for ethanol sensing. *Sens Actuat B Chem* 146: 559–65.
52. Wilson AD (2013) Diverse applications of electronic-nose technologies in agriculture and forestry. *Sensors* 13: 2295–348.
53. Cole MD (2003) *The analysis of controlled substances*. John Wiley & Sons Ltd., Chichester, UK.
54. Groombridge CJ (1996) NMR spectroscopy in forensic science. *Annu Rep NMR Spectrosc* 32: 215–97.
55. Bell SEJ, Burns DT, Dennis AC, Matchett LJ, Speers JS (2000) Composition profiling of seized ecstasy tablets by Raman Spectroscopy. *Analyst* 125: 1811–5.
56. Moore JM, Casale JF (1998) Cocaine profiling methodology-recent advances. *Forensic Sci Rev* 10: 13–46.
57. Phillips SA, Doyle S, Philp L, Coleman M (2003) Proceedings network developing forensic applications of stable isotope ratio mass spectrometry conference 2002. *Sci Justice* 43: 153–60.
58. Pagano B, Lauri I, De Tito S, Persico G, Chini MG, et al. (2013) Use of NMR in profiling of cocaine seizures. *Forensic Sci Int* 231: 120–4.
59. He J-L, Wu Z-S, Zhou H, Wang H-Q, Jiang J-H, et al. (2010) Fluorescence aptameric sensor for strand displacement amplification detection of cocaine. *Anal Chem* 82: 1358–64.
60. Haddi Z, Amari A, Alami H, El Bari N, Llobet E, et al. (2011) A portable electronic nose system for the identification of cannabis-based drugs. *Sens Actuat B* 155: 456–63.
61. Rendle DF, Taylor JF (1997) Application of XRF to the detection and estimation of metals in toxicological specimens. *Adv X-ray Anal* 39: 869–80.
62. Wilson AD (2014) Identification of insecticide residues with a conducting-polymer electronic nose. *Chem Sensors* 4: 1–10.
63. Wilson AD (2013) Fungicide residue identification and discrimination using a conducting polymer electronic-nose. In: *Proceedings of the Fourth International Conference on Sensor Device Technologies and Applications*, Barcelona, Spain, Xpert Publishing Services, Wilmington, DE, USA.
64. Urdea M, Penny LA, Olmsted SS, Giovanni MY, Kaspar P, et al. (2006) Requirements for high impact diagnostics in the developing world. *Nature* 444: 73–9.
65. Wilson AD, Lester DG, Oberle CS (2004) Development of conductive polymer analysis for the rapid detection and identification of phytopathogenic microbes. *Phytopathology* 94: 419–31.
66. Vass AA, Smith RR, Thompson CV, Burnett MN, Wolf DA, et al. (2004) Decompositional odor analysis database. *J Forensic Sci* 49: 760–9.
67. Vass AA (2008) Review of soil analysis in forensic taphonomy: Chemical and biological effects of buried human remains. *J Forensic Sci* 53: 1484–5.
68. Vass AA (2010) Dust to Dust - how a human body decomposes. *Scientific American Special Issue: The End*, 56–59.
69. Becher C, Kaul P, Mitrovics J, Warmer J (2010) The detection of evaporating hazardous material released from moving sources using a gas sensor network. *Sens Actuat B Chem* 146: 513–20.
70. Lamagna A, Reich S, Rodríguez D, Boselli A, Cicerone D (2008) The use of an electronic nose to characterize emissions from a highly polluted river. *Sens Actuat B Chem* 131: 121–4.
71. Fuchs S, Strobel P, Siadat M, Lumbreras M (2008) Evaluation of unpleasant odor with a portable electronic nose. *Mater Sci Eng C* 28: 949–53.
72. Tillman ES, Koscho ME, Grubbs RH, Lewis NS (2003) Enhanced sensitivity to and classification of volatile carboxylic acids using arrays of linear poly(ethylenimine)-carbon black composite vapor detectors. *Anal Chem* 75: 1748–53.
73. Bondy PK, Rosenberg LE (1980) *Metabolic Control and Disease*, 8th ed.; W.B. Sanders: Philadelphia, PA, USA.
74. Milne MD (1964) Disorders of amino-acid transport. *Brit Med J* 1: 327–36.
75. Vass AA, Smith RR, Thompson CV, Burnett MN, Dulgerian N, et al. (2008) Odor analysis of decomposing buried human remains. *J Forensic Sci* 53: 384–91.
76. Parkinson RA, Dias KR, Horswell J, Greenwood P, Banning N, et al. (2009) *Microbial community analysis of human decomposition in soil*. Criminal and Environmental Soil Forensics, Springer, New York.
77. Larson DO, Vass AA, Wise M (2011) Advanced scientific methods and procedures in the forensic investigation of clandestine graves. *J Contemp Crim Just* 27: 149–82.
78. Brogan KL, Walt DR (2005) Optical fiber-based sensors: application to chemical biology. *Curr Opin Chem Biol* 9: 494–500.

79. Gao T, Tillman ES, Lewis NS (2005) Detection and classification of volatile organic amines and carboxylic acids using arrays of carbon black-dendrimer composite vapor detectors. *Chem Mater* 17: 2904-11.
80. Tillman ES, Lewis NS (2003) Mechanism of enhanced sensitivity of linear poly-(ethylenimine)-carbon black composite detectors to carboxylic acid vapors. *Sens Actuatur B Chem* 96: 329-42.
81. Huo R, Agapiou A, Bocos-Bintintan V, Brown LJ, Burns C, et al. (2011) The trapped human experiment. *J Breath Res* 5: 046006.
82. Vautz W, Slodzynski R, Hariharan C, Seifert L, Nolte J, et al. (2013) Detection of metabolites of trapped humans using ion mobility spectrometry coupled with gas chromatography. *Anal Chem* 85: 2135-42.
83. Montgomery JA, Jette M, Huot S, des Rosiers C (1993) Acyloloin production from aldehydes in the perfused rat heart: the potential role of pyruvate dehydrogenase. *Biochem J* 294: 727-33.
84. Keller JN, Pang Z, Geddes JW, Begley JO, Germeyer A, et al. (1997) Impairment of glucose and glutamate transport and induction of mitochondrial oxidative stress and dysfunction in synaptosomes by amyloid β -peptide: role of the lipid peroxidation product 4-hydroxynonenal. *J Neurochem* 69: 273-84.
85. Vivanti A, Harvey K, Ash S (2010) Developing a quick and practical screen to improve the identification of poor hydration in geriatric and rehabilitative care. *Arch Gerontol Geriatr* 50: 156-64.
86. Likhodii SS, Musa K, Cunnane SC (2002) Breath acetone as a measure of systemic ketosis assessed in a rat model of the ketogenic diet. *Clin Chem* 48: 115-20.
87. Statheropoulos M, Agapiou A, Georgiadou A (2006) Analysis of expired air of fasting male monks at Mount Athos. *J Chromatography B* 832: 274-9.
88. Rimeika R, Čiplys D, Poderys V, Rotomskis R, Shur MS (2009) Fast-response surface acoustic wave humidity sensor based on hematoporphyrin film. *Sens Actuatur B Chem* 137: 592-6.
89. Dini F, Capuano R, Strand T, Ek A-C, Lindgren M, et al. (2013) Volatile emissions from compressed tissue. *Plos One* 8: 1-9.89
90. Gallagher M, Wysocki CJ, Leyden JJ, Spielman AI, Sun X, et al. (2008) Analyses of volatile organic compounds from human skin. *Br J Dermatol* 159: 780-91.
91. Jha PK, Sawicka KM, Gouma PI (2004) Nanocomposite materials for electronic nose. International Symposium of Research Students on Materials Science and Engineering, Indian Institute of Technology Madras, Chennai, India.
92. Littarru P (2007) Environmental odour assessment from waste treatment plants: Dynamic olfactometry in combination with sensorial analysers "electronic noses". *Waste Manage* 27: 302-9.
93. Mochalski P, King J, Klieber M, Unterkofler K, Hinterhuber H, et al. (2013) Blood and breath levels of selected volatile organic compounds in healthy volunteers. *Analyst* 138: 2134-45.
94. Soso SB, Koziel JA, Johnson A, Lee YJ, Fairbanks WS (2014) Analytical methods for chemical and sensory characterization of scent-markings in large wild mammals: A review. *Sensors* 14: 4428-65.
95. Forbes SL, Perrault KA (2014) Decomposition odour profiling in the air and soil surrounding vertebrate carrion. *Plos One* 9: e95107.
96. Phillips M, Cataneo RN, Chaturvedi A, Kaplan PD, Libardoni M, et al. (2013) Detection of an extended human volatome with comprehensive two-dimensional gas chromatography time-of-flight mass spectrometry. *Plos One* 8: e75274.
97. Bos LDJ, Sterk PJ, Schultz MJ (2013) Volatile metabolites of pathogens: A systemic review. *Plos Pathogens* 9: e1003311.
98. Netzer M, Millonig G, Osl M, Pfeifer B, Praun S, et al. (2009) A new ensemble-based algorithm for identifying breath gas marker candidates in liver disease using ion molecule reaction mass spectrometry. *Bioinformatics* 25: 941-7.
99. Lett eron P, Duchatelle V, Berson A, Fromenty B, Fisch C, et al. (1993) Increased ethane exhalation, an in vivo index of lipid peroxidation, in alcohol-abusers. *Gut* 34: 409-14.
100. Wlodzimirow KA, Abu-Hanna A, Schultz MJ, Maas MAW, Bos LDJ, et al. (2014) Exhaled breath analysis with electronic nose technology for detection of acute liver failure in rats. *Biosens Bioelectron* 53: 129-34.
101. Scholpp J, Schubert JK, Miekisch W, Geiger K (2002) Breath markers and soluble lipid peroxidation markers in critically ill patients. *Clin Chem Lab Med* 40: 587-94.
102. Schubert JK, Muller WP, Benzing A, Geiger K (1998) Application of a new method for analysis of exhaled gas in critically ill patients. *Intensive Care Med* 24: 415-21.
103. Chambers ST, Syhre M, Murdoch DR, McCartin F, Epton MJ (2009) Detection of 2-pentylfuran in the breath of patients with *Aspergillus fumigatus*. *Med Mycol* 47: 468-76.
104. de Heer K, van der Schee MP, Zwinderman K, van den Berk IAH, Visser CE, et al. (2013) Electronic nose technology for detection of invasive pulmonary aspergillosis in prolonged chemotherapy-induced neutropenia: a proof-of-principle study. *J Clin Microbiol* 51: 1490-5.
105. Olopade CO, Zakkar M, Swedler WI, Rubinstein I (1997) Exhaled pentane levels in acute asthma. *Chest* 111: 862-5.
106. Paredi P, Kharitonov SA, Barnes PJ (2000) Elevation of exhaled ethane concentration in asthma. *Am J Respir Crit Care Med* 162: 1450-4.
107. Larstad MA, Toren K, Bake B, Olin AC (2007) Determination of ethane, pentane and isoprene in exhaled air—effects of breath-holding, flow rate and purified air. *Acta Physiol (Oxf)* 189: 87-98.
108. Kostikas K, Papatheodorou G, Psathakis K, Panagou P, Loukides S (2003) Prostaglandin E2 in the expired breath condensate of patients with asthma. *Eur Respir J* 22: 743-7.
109. Kostikas K, Gaga M, Papatheodorou G, Karamanis T, Orphanidou D, et al. (2005) Leukotriene B4 in exhaled breath condensate and sputum supernatant in patients with COPD and asthma. *Chest* 127: 1553-9.
110. Montuschi P, Corradi M, Ciabattini G, Nightingale J, Kharitonov SA, et al. (1999) Increased 8-isoprostane, a marker of oxidative stress, in exhaled condensate of asthma patients. *Am J Respir Crit Care Med* 160: 216-20.
111. Smith AD, Cowan JO, Filsell S, McLachlan C, Monti-Sheehan G, et al. (2004) Diagnosing asthma: Comparisons between exhaled nitric oxide measurements and conventional tests. *Am J Respir Crit Care Med* 169: 473-8.

112. Dragonieri S, Schot R, Mertens B, Le Cessie S, Gauw S, et al. (2007) An electronic nose in the discrimination of patients with asthma and controls. *J Allergy Clin Immunol* 120: 856-62.
113. Caldeira M, Barros AS, Bilelo MJ, Parada A, Camara JS, et al. (2011) Profiling allergic asthma volatile metabolic patterns using a headspace-solid phase microextraction/gas chromatography based methodology. *J Chromatogr A* 1218: 3771-80.
114. Caldeira M, Perestrelo R, Barros AS, Bilelo MJ, Morete A, et al. (2012) Allergic asthma exhaled breath metabolome: A challenge for comprehensive two-dimensional gas chromatography. *J Chromatogr A* 1254: 87-97.
115. Ibrahim B, Basanta M, Cadden P, Singh D, Douce D, et al. (2011) Non-invasive phenotyping using exhaled volatile organic compounds in asthma. *Thorax* 66: 804-9.
116. Fens N, Zwinderman AH, van der Schee MP, de Nijs SB, Dijkers E, et al. (2009) Exhaled breath profiling enables discrimination of chronic obstructive pulmonary disease and asthma. *Am J Respir Crit Care Med* 180: 1076-82.
117. Mitra AP, Datar RH, Cote RJ (2006) Molecular pathways in invasive bladder cancer: new insights into mechanisms, progression, and target identification. *J Clin Oncol* 24: 5552-64.
118. Phillips M, Cataneo RN, Ditkoff BA, Fisher P, Greenberg J, et al. (2003) Volatiles markers of breast cancer in the breath. *The Breast J* 9: 184-91.
119. Hakim M, Billan S, Tisch U, Peng G, Dvorkind I, et al. (2011) Diagnosis of head-and-neck cancer from exhaled breath. *Br J Cancer* 104: 1649-55.
120. Shirasu M, Nagai S, Hayashi R, Ochiai A, Touhara K (2009) Dimethyl trisulfide as a characteristic odor associated with fungating cancer wounds. *Biosci Biotechnol Biochem* 73: 2117-20.
121. Phillips M, Gleeson K, Huges JMB, Greenberg J, Cataneo RN, et al. (1999) Volatile organic compounds in breath as markers of lung cancer: a cross-sectional study. *Lancet* 353: 1930-3.
122. Phillips M, Cataneo RN, Cummin AR, Gagliardi AJ, Gleeson K, et al. (2003) Detection of lung cancer with volatile markers in the breath. *Chest* 123: 2115-23.
123. Di Natale C, Macagnano A, Martinelli E, Paolesse R, D'Arcangelo G, et al. (2003) Lung cancer identification by the analysis of breath by means of an array of non-selective gas sensors. *Biosens Bioelectron* 18: 1209-18.
124. Preti G, Labows JN, Kostelc JG, Aldinger S, Daniele R (1988) Analysis of lung air from patients with bronchogenic carcinoma and controls using gas chromatography-mass spectrometry. *J Chromatogr* 432: 1-11.
125. Fuchs P, Loeseke C, Schubert JK, Miekisch W (2010) Breath gas aldehydes as biomarkers of lung cancer. *Int J Cancer* 126: 2663-70.
126. Poli D, Goldoni M, Corradi M, Acampa O, Carboganni P, et al. (2010) Determination of aldehydes in exhaled breath of patients with lung cancer by means of on-fiber-derivatization SPME-GC/MS. *J Chromatogr B Analyt Technol Biomed Life Sci* 878: 2643-51.
127. Song G, Qin T, Liu H, Xu GB, Pan YY, et al. (2010) Quantitative breath analysis of volatile organic compounds of lung cancer patients. *Lung Cancer* 67: 227-31.
128. Kischkel S, Miekisch W, Sawacki A, Straker EM, Trefz P, et al. (2010) Breath biomarkers for lung cancer detection and assessment of smoking related effects—confounding variables, influence of normalization and statistical algorithms. *Clin Chim Acta* 411: 1637-44.
129. Skeldon KD, McMillan LC, Wyse CA, Monk SD, Gibson G, et al. (2006) Application of laser spectroscopy for measurement of exhaled ethane in patients with lung cancer. *Respir Med* 100: 300-6.
130. Bajtarevic A, Ager C, Pienz M, Klieber M, Schwarz K, et al. (2009) Noninvasive detection of lung cancer by analysis of exhaled breath. *BMC Cancer* 9: 348-64.
131. Crohns M, Saarelainen S, Laitinen J, Peltonen K, Alho H, et al. (2009) Exhaled pentane as a possible marker for survival and lipid peroxidation during radiotherapy for lung cancer—a pilot study. *Free Radic Res* 43: 965-74.
132. Peled N, Hakim M, Bunn Jr. PA, Miller YE, Kennedy TC, et al. (2012) Non-invasive breath analysis of pulmonary nodules. *J Thorac Oncol* 7: 1528-33.
133. Dragonieri S, Annema JT, Schot R, van der Schee MP, Spanevello A, et al. (2009) An electronic nose in the discrimination of patients with non-small cell lung cancer and COPD. *Lung Cancer* 64: 166-70.
134. D'Amico A, Pennazza G, Santonico M, Martinelli E, Roscioni C, et al. (2010) An investigation on electronic nose diagnosis of lung cancer. *Lung Cancer* 68: 170-6.
135. Gaspar EM, Lucena AF, Duro da Costa J, Chaves das Neves H (2009) Organic metabolites in exhaled human breath—a multivariate approach for identification of biomarkers in lung disorders. *J Chromatogr A* 1216: 2749-56.
136. Kischkel S, Miekisch W, Sawacki A, Straker EM, Trefz P, et al. (2010) Breath biomarkers for lung cancer detection and assessment of smoking related effects—confounding variables, influence of normalization and statistical algorithms. *Clin Chim Acta* 411: 1637-44.
137. Ligor M, Ligor T, Bajtarevic A, Ager C, Pienz M, et al. (2009) Determination of volatile organic compounds in exhaled breath of patients with lung cancer using solid phase microextraction and gas chromatography mass spectrometry. *Clin Chem Lab Med* 47: 550-60.
138. Peng G, Tisch U, Adams O, Hakim M, Shehada N, et al. (2009) Diagnosing lung cancer in exhaled breath using gold nanoparticles. *Nat Nanotechnol* 4: 669-73.
139. Peng G, Hakim M, Broza YY, Billan S, Abdah-Bortnyak R, et al. (2010) Detection of lung, breast, colorectal, and prostate cancers from exhaled breath using a single array of nanosensors. *BJC* 103: 542-51.
140. Phillips M, Altorki N, Austin JH, Cameron RB, Cataneo RN, et al. (2007) Prediction of lung cancer using volatile biomarkers in breath. *Cancer Biomark* 3: 95-109.
141. Phillips M, Altorki N, Austin JH, Cameron RB, Cataneo RN, et al. (2008) Detection of lung cancer using weighted digital analysis of breath biomarkers. *Clin Chim Acta* 393: 76-84.
142. Poli D, Goldoni M, Caglieri A, Ceresa G, Acampa O, et al. (2008) Breath analysis in non small cell lung cancer patients after surgical tumour resection. *Acta Biomed* 79: 64-72.
143. Rudnicka J, Kowalkowski T, Ligor T, Buszewski B (2011) Determination of volatile organic compounds as biomarkers of lung cancer by SPME-GC-TOF/MS and chemometrics. *J Chromatogr B Analyt Technol Biomed Life Sci* 879: 3360-6.

144. Ulanowska A, Kowalkowski T, Trawinska E, Buszewski B (2011) The application of statistical methods using VOCs to identify patients with lung cancer. *J Breath Res* 5: 046008.
145. Machado RF, Laskowski D, Deffenderfer O, Burch T, Zheng S, et al. (2005) Detection of lung cancer by sensor array analyses of exhaled breath. *Am J Respir Crit Care Med* 171: 1286-91.
146. Machado RF (2009) Identifying chronic obstructive pulmonary disease and asthma by exhaled breath analysis: Does the e-nose know? *Am J Respir Crit Care Med* 180: 1038-9.
147. Poli D, Carbognani P, Corradi M, Goldoni M, Acampa O, et al. (2005) Exhaled volatile organic compounds in patients with non-small cell lung cancer: Cross sectional and nested short-term follow-up study. *Respir Res* 6: 71.
148. Mazzone PJ, Wang X-F, Xu Y, Mekhail T, Beukemann MC, et al. (2012) Exhaled breath analysis with a colorimetric sensor array for the identification and characterization of lung cancer. *J Thorac Oncol* 7: 137-42.
149. Kwak J, Gallagher M, Ozdener MH, Wysocki CJ, Goldsmith BR, et al. (2013) Volatile biomarkers from human melanoma cells. *J Chromatogr B* 931: 90-6.
150. Kaji H, Hisamura M, Saito N, Muraio M (1978) Gas chromatographic determination of volatile sulphur compounds in expired alveolar air in hepatopathic patients. *J Chromatogr* 145: 464-8.
151. Balint B, Kharitonov SA, Hanazawa T, Donnelly LE, Shah PL, et al. (2001) Increased nitrotyrosine in exhaled breath condensate in cystic fibrosis. *Eur Respir J* 17: 1201-7.
152. Borrill ZL, Starkey RC, Singh SD (2007) Variability of exhaled breath condensate leukotriene B4 and 8-isoprostane in COPD patients. *Int J Chron Obstruct Pulmon Dis* 2: 71-6.
153. van Beurden WJ, Harff GA, Dekhuijzen PNR, van den Bosch MJ, Creemers JP, et al. (2002) An efficient and reproducible method for measuring hydrogen peroxide in exhaled breath condensate. *Respir Med* 96: 197-203.
154. van Beurden WJ, Dekhuijzen PN, Harff GA, Smeenk FW (2002) Variability of exhaled hydrogen peroxide in stable COPD patients and matched healthy controls. *Respiration* 69: 211-6.
155. Corradi M, Pesci A, Casana R, Alinovi R, et al. (2003) Nitrate in exhaled breath condensate of patients with different airway diseases. *Nitric Oxide - Biol Chem* 8: 26-30.
156. Corradi M, Majori M, Cacciani GC, Consigli GF, de'Munari E, et al. (1999) Increased exhaled nitric oxide in patients with stable chronic obstructive pulmonary disease. *Thorax* 54: 572-5.
157. Paredi P, Kharitonov SA, Leak D, Ward S, Cramer D, et al. (2000) Exhaled ethane, a marker of lipid peroxidation, is elevated in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 162: 369-73.
158. Fens N, Roldaan AC, van der Schee MP, Boksem RJ, Zwinderman AH, et al. (2011) External validation of exhaled breath profiling using an electronic nose in the discrimination of asthma with fixed airways obstruction and chronic obstructive pulmonary disease. *Clin Exp Allergy* 41: 1371-8.
159. Hattesoehl AD, Jorres RA, Dressel H, Schmid S, Vogelmeier C, et al. (2011) Discrimination between COPD patients with and without alpha 1-antitrypsin deficiency using an electronic nose. *Respirology* 16: 1258-64.
160. Hauschild AC, Baumbach JJ, Baumbach J (2012) Integrated statistical learning of metabolic ion mobility spectrometry profiles for pulmonary disease identification. *Genet Mol Res* 11: 2733-44.
161. Phillips CO, Syed Y, Parthala NM, Zwiggelaar R, Claypole TC, et al. (2012) Machine learning methods on exhaled volatile organic compounds for distinguishing COPD patients from healthy controls. *J Breath Res* 6: 036003.
162. Timms C, Thomas PS, Yates DH (2012) Detection of gastro-oesophageal reflux disease (GORD) in patients with obstructive lung disease using exhaled breath profiling. *J Breath Res* 6: 016003
163. Fens N, de Nijs SB, Peters S, Dekker T, Knobel HH, et al. (2011) Exhaled air molecular profiling in relation to inflammatory subtype and activity in COPD. *Eur Respir J* 38: 1301-9.
164. Incalzi RA, Scarlata S, Pennazza G, Santonico M, Pedone C (2014) Chronic obstructive pulmonary disease in the elderly. *Eur J Intern Med* 25: 320-28.
165. Skrupskii VA (1995) Gas chromatographic analysis of ethanol and acetone in the air exhaled by patients. *Klin Lab Diagn* 4: 35-8.
166. Carpagnano GE, Barnes PJ, Geddes DM, Hodson ME, Kharitonov SA (2003) Increased leukotriene B4 and interleukin-6 in exhaled breath condensate in cystic fibrosis. *Am J Respir Crit Care Med* 167: 1109-12.
167. Barker M, Hengst M, Schmid J, Buers HJ, Mittermaier B, et al. (2006) Volatile organic compounds in the exhaled breath of young patients with cystic fibrosis. *Eur Respir J* 27: 929-36.
168. Kamboures MA, Blake DR, Cooper DM, Newcomb RL, Barker M, et al. (2005) Breath sulfides and pulmonary function in cystic fibrosis. *Proc Natl Acad Sci USA* 102: 15762-7.
169. Shestivska V, Nemeš A, Drevinek P, Sovova K, Dryahina K, et al. (2011) Quantification of methyl thiocyanate in the headspace of *Pseudomonas aeruginosa* cultures and in the breath of cystic fibrosis patients by selected ion flow tube mass spectrometry. *Rapid Commun Mass Spectrom* 25:2459-67.
170. Paredi P, Kharitonov SA, Leak D, Shah PL, Cramer D, et al. (2000) Exhaled ethane is elevated in cystic fibrosis and correlates with carbon monoxide levels and airway obstruction. *Amer J Respir Crit Care Med* 161: 1247-51.
171. McGrath LT, Patrick R, Mallon P, Dowey L, Silke B, et al. (2000) Breath isoprene during acute respiratory exacerbation in cystic fibrosis. *Eur Respir J* 16: 1065-9.
172. Enderby B, Smith D, Carroll W, Lenney W (2009) Hydrogen cyanide as a biomarker for *Pseudomonas aeruginosa* in the breath of children with cystic fibrosis. *Pediatr Pulmonol* 44: 142-7.
173. Gilchrist FJ, Razavi C, Webb AK, Jones AM, Spanel P, et al. (2012) An investigation of suitable bag materials for the collection and storage of breath samples containing hydrogen cyanide. *J Breath Res* 6:036004.
174. Robroeks CM, van Berkel JJ, Dallinga JW, Jöbsis Q, Zimmermann LJ, et al. (2010) Metabolomics of volatile organic compounds in cystic fibrosis patients and controls. *Pediatr Res* 68: 75-80.
175. Paff T, van der Schee MP, Daniels JMA, Pals G, Postmus PE, et al. (2013) Exhaled molecular profiles in the assessment of cystic fibrosis and primary ciliary dyskinesia. *J Cystic Fibros* 12: 454-60.

176. Ping W, Yi P, Haibao X, Farange S (1997) A novel method for diabetes diagnosis based on electronic nose. *Biosens Bioelectron* 12: 1031-36.
177. Novak BJ, Blake DR, Meinardi S, Rowland FS, Pontello A, et al. (2007) Exhaled methyl nitrate as a noninvasive marker of hyperglycemia in type 1 diabetes. *Proc Nat Acad Sci* 104: 15613-18.
178. Cristescu SM, Gietema HA, Blanchet L, Kruitwagen CL, Munnik P, et al. (2011) Screening for emphysema via exhaled volatile organic compounds. *J Breath Res* 5:046009.
179. Van den Velde S, van Steenberghe D, Van Hee P, Quirynen M (2009) Detection of odorous compounds in breath. *J Dent Res* 88: 285-9.
180. Okell CC, Elliott SD (1935) Bacteriaemia and oral sepsis with special reference to the aetiology of subacute endocarditis. *The Lancet* 2: 869-72.
181. Drangsholt M T (1998) A new causal model of dental diseases associated with endocarditis. *Ann Periodontol* 3: 184-96.
182. Lacassin F, Hoen B, Lepout C, Selton-Suty C, Delahaye F, et al. (1995) Procedures associated with infective endocarditis in adults: a case-control study. *Eur Heart J* 16: 1968-74.
183. Tangerman A, Meuwese-Arends MT, Jansen JB (1994) Cause and composition of foetor hepaticus. *Lancet* 343: 483.
184. Chen S, Mahadevan V, Zieve L (1970) Volatile fatty acids in the breath of patients with cirrhosis of the liver. *J Lab Clin Med* 75: 622-7.
185. Van den Velde S, Nevens F, Van Hee P, van Steenberghe D, Quirynen M (2008) GC-MS analysis of breath odor compounds in liver patients. *J Chromatography B* 875: 344-8.
186. Kaji H, Hisamura M, Sato N, Murao M (1978) Evaluation of volatile sulfur compounds in the expired alveolar gas in patients with liver cirrhosis. *Clin Chim Acta* 85: 279-84.
187. Hisamura M (1979) Quantitative analysis of methyl mercaptan and dimethyl sulfide in human expired alveolar gas and its clinical application: Study in normal subjects and patients with liver diseases. *Nippon Naika Gakkai Zasshi* 68: 1284-92.
188. Lee J, Ngo J, Blake D, Meinardi S, Pontello AM, et al. (2009) Improved predictive models for plasma glucose estimation from multi-linear regression analysis of exhaled volatile organic compounds. *J Appl Physiol* 107: 155-60.
189. Ondrula D, Nelson RL, Andrianopoulos G, Schwartz D, Abcarian H, et al. (1993) Quantitative determination of pentane in exhaled air correlates with colonic inflammation in the rat colitis model. *Dis Colon Rectum* 36: 457-62.
190. Kokoszka J, Nelson RL, Swedler WI, Skosey J, Abcarian H (1993) Determination of inflammatory bowel disease activity by breath pentane analysis. *Dis Colon Rectum* 36: 597-601.
191. Sedghi S, Keshavarzian A, Klamut M, Eiznhamer D, Zarling EJ (1994) Elevated breath ethane levels in active ulcerative colitis: evidence for excessive lipid peroxidation. *Am J Gastroenterol* 89: 2217-21.
192. Pelli MA, Trovarelli G, Capodicasa E, De Medio GE, Bassotti G (1999) Breath alkanes determination in ulcerative colitis and Crohn's disease. *Dis Colon Rectum* 42: 71-6.
193. Dobbelaar P, Mottram TT, Nyabadza C, Hobbs PJ, Elliott-Martin RJ, et al. (1996) Detection of ketosis in dairy cows by analysis of exhaled breath. *Vet Quart* 18: 151-2.
194. Kanoh S, Kobayashi H, Motoyoshi K (2005) Exhaled ethane: an in vivo biomarker of lipid peroxidation in interstitial lung diseases. *Chest* 128: 2387-92.
195. Dragonieri S, Brinkman P, Mouw E, Zwinderman AH, Carratú P, et al. (2013) An electronic nose discriminates exhaled breath of patients with untreated pulmonary sarcoidosis from controls. *Respir Med* 107: 1073-8.
196. Chapman EA, Thomas PS, Stone E, Lewis C, Yates DH (2011) A breath test for malignant mesothelioma using an electronic nose. *Eur Respir J* 40: 448-54.
197. Dragonieri S, van der Schee MP, Massaro T, Schiavulli N, Brinkman P, et al. (2012) An electronic nose distinguishes exhaled breath of patients with malignant pleural mesothelioma from controls. *Lung Cancer* 75: 326-31.
198. de Gennaro G, Dragonieri S, Longobardi F, Musti M, Stallone G, et al. (2010) Chemical characterization of exhaled breath to differentiate between patients with malignant pleural mesothelioma from subjects with similar professional asbestos exposure. *Anal Bioanal Chem* 398: 3043-50.
199. Hakim M, Broza YY, Barash O, Peled N, Phillips M, et al. (2012) Volatile organic compounds of lung cancer and possible biochemical pathways. *Chem Rev* 112: 5949-66.
200. Wehinger A, Schmid A, Mechtcheriakov S, Ledochowski M, Grabmer C, et al. (2007) Lung cancer detection by proton transfer reaction mass-spectrometric analysis of human breath gas. *Int J Mass Spectrom* 265: 49-59.
201. Voss A, Baier V, Reisch R, Von Roda K, Elsner P, et al. (2005) Smelling renal dysfunction via electronic nose. *Ann Biomed Eng* 33: 656-60.
202. Hanson CW, Steinberger HA (1998) The use of a novel electronic nose to diagnose the presence of intrapulmonary infection. *Anesthesiology* 87: A269.
203. Humad S, Zarling E, Clapper M, Skosey JL (1988) Breath pentane excretion as a marker of disease activity in rheumatoid arthritis. *Free Radic Res Commun* 5: 101-6.
204. Smith K, Sines J (1960) Demonstration of a peculiar odor in the sweat of schizophrenic patients. *AMA Arch Gen Psych* 2: 184-8.
205. Smith K, Thompson GF, Koster HD (1969) Sweat in schizophrenic patients: Identification of the odorous substance. *Science* 166: 398-9.
206. Phillips M, Sabas M, Greenberg J (1993) Increased pentane and carbon disulphide in the breath of patients with schizophrenia. *J Clin Pathol* 46: 861-4.
207. Sidbury JB, Smith EK, Harlan W (1967) An inborn error of short-chain fatty acid metabolism: The odor of sweaty feet syndrome. *J Pediatr* 70: 8-15.
208. Syhre M, Manning L, Phuanukoonnon S, Harino P, Chambers ST (2009) The scent of *Mycobacterium tuberculosis*-part II breath. *Tuberculosis* 89: 263-6.
209. Phillips M, Cataneo RN, Condos R, Ring Erickson GA, Greenberg J, et al. (2007) Volatile biomarkers of pulmonary tuberculosis in the breath. *Tuberculosis* 87: 44-52.
210. Phillips M, Basa-Dalay V, Bothamley G, Cataneo RN, Lam PK, et al. (2010) Breath biomarkers of active pulmonary tuberculosis. *Tuberculosis* 90: 145-51.
211. Phillips M, Basa-Dalay V, Blais J, Bothamley G, Chaturvedi A, et al. (2012) Point-of-care breath test for biomarkers of active pulmonary tuberculosis. *Tuberculosis* 92: 314-20.2
212. Bruins M, Rahim Z, Bos A, van de Sande WWJ, et al. (2013) Diagnosis of active tuberculosis by e-nose analysis of exhaled air. *Tuberculosis* 93: 232-8.
213. du Preez I, Loots DT (2013) New sputum metabolite markers implicating adaptations of the host to *Mycobacterium tuberculosis*, and vice versa. *Tuberculosis* 93: 330-7.

214. Filipiak W, Sponring A, Baur MM, Ager C, Filipiak A, et al. (2012) Characterization of volatile metabolites taken up by or released from *Streptococcus pneumoniae* and *Haemophilus influenzae* by using GC-MS. *Microbiology* 158: 3044–53.
215. Hanson CW, Thaler ER (2005) Electronic nose prediction of a clinical pneumonia score: biosensors and microbes. *Anesthesiology* 102: 63–8.
216. Hockstein NG, Thaler ER, Torigian D, Miller WT Jr, Deffenderfer O, et al. (2004) Diagnosis of pneumonia with an electronic nose: correlation of vapor signature with chest computed tomography scan findings. *Laryngoscope* 114: 1701–5.
217. Hockstein NG, Thaler ER, Lin Y, Lee DD, Hanson CW (2005) Correlation of pneumonia score with electronic nose signature: A prospective study. *Ann Otol Rhinol Laryngol* 114: 504–8.
218. Filipiak W, Sponring A, Baur MM, Filipiak A, Ager C (2012) Molecular analysis of volatile metabolites released specifically by *Staphylococcus aureus* and *Pseudomonas aeruginosa*. *BMC Microbiol* 12: 113.
219. Boots AW, van Berkel JJBN, Dallinga JW, Smolinska A, Wouters EF, et al. (2012) The versatile use of exhaled volatile organic compounds in human health and disease. *J Breath Res* 6: 027108.
220. Montuschi P, Mores N, Trové A, Mondino C, Barnes PJ (2013) The electronic nose in respiratory medicine. *Respiration* 85: 72–84.
221. Biller H, Holz O, Windt H, Koch W, Müller M, et al (2011) Breath profiles by electronic nose correlate with systemic markers but not ozone response. *Respir Med* 105: 1352–63.
222. Prabhakar A, Iglesias RA, Shan X, Xian X, Zhang L, et al. (2012) Online sample conditioning for portable breath analyzers. *Anal Chem* 84: 7172–8.
223. Konvalina G, Haick H (2014) Sensors for breath testing: From nanomaterials to comprehensive disease detection. *Acc Chem Res* 47: 66–76.
224. Mochalski P, Sponring A, King J, Unterkofler K, Troppmair J, et al. (2013) Release and uptake of volatile organic compounds by human hepatocellular carcinoma cells (HepG2) in vitro. *Cancer Cell Internat* 13: 72–81.
225. Amal H, Ding L, Liu BB, Tisch U, Xu ZQ, et al. (2012) The scent fingerprint of hepatocarcinoma: in-vitro metastasis prediction with volatile organic compounds (VOCs). *Int J Nanomedicine* 7: 4135–46.
226. Pennazza G, Santonico M, Martinelli E, Paolesse R, Tamburrelli V, et al. (2011) Monitoring of melanoma released volatile compounds by a gas sensors array: From in vitro to in vivo experiments. *Sens Actuat B* 154: 288–94.
227. Righettoni M, Tricoli A, Gass S, Schmid A, Aman A (2012) Breath acetone monitoring by portable Si:WO₃ gas sensors. *Anal Chim Acta* 738: 69–75.
228. Rogers PH, Benkstein KD, Semancik S (2012) Machine learning applied to chemical analysis: Sensing multiple biomarkers in simulated breath using a temperature-pulsed electronic-nose. *Anal Chem* 84: 9774–81.
229. Barash O, Peled N, Tisch U, Bunn PA, Hirsch FR, et al. (2012) Classification of lung cancer histology by gold nanoparticle sensors. *Nanomedicine* 8: 580–9.
230. Broza YY, Kremer R, Tisch U, Gevorkyan A, Shibani A, et al. (2013) A nanomaterial-based breath test for short-term follow-up after lung tumor resection. *Nanomedicine* 9: 15–21.
231. Tisch U, Billan S, Ilouze M, Phillips M, Peled N, et al. (2012) Volatile organic compounds in exhaled breath as biomarkers for the early detection and screening of lung cancer. *CML Lung Cancer* 5: 107–17.
232. Xu ZQ, Broza YY, Ionsecu R, Tisch U, Ding L, et al. (2013) A nanomaterial-based breath test for distinguishing gastric cancer from benign gastric conditions. *British J Cancer* 108: 941–50.
233. Španěl P, Smith D (2011) Volatile compounds in health and disease. *Curr Opin Clin Nutr Metabol Care* 14: 455–60.
234. Michael R, Thorn S, Greenman J (2012) Microbial volatile compounds in health and disease conditions. *J Breath Res* 6: 024001.
235. Fuchs D, Jamnig H, Heininger P, Klieber M, Schroecksnadel S, et al. (2012) Decline of exhaled isoprene in lung cancer patients correlates with immune activation. *J Breath Res* 6: 027101.
236. Španěl P, Dryahina K, Smith D (2013) A quantitative study of the influence of inhaled compounds on their concentrations in exhaled breath. *J Breath Res* 7: 017106.
237. Filipiak W, Ruzsanyi V, Mochalski P, Filipiak A, Bajtarevic A, et al. (2012) Dependence of exhaled breath composition on exogenous factors, smoking habits and exposure to air pollutants. *J Breath Res* 6: 036008.
238. Filipiak W, Filipiak A, Ager C, Wiesenhofer H, Amann A (2012) Optimization of sampling parameters for collection and preconcentration of alveolar air by needle traps. *J Breath Res* 6: 027107.
239. Mochalski P, King J, Unterkofler K, Amann A (2013) Stability of selected volatile breath constituents in tedlar, kynar and flexfilm sampling bags. *Analyst* 138: 1405–18.
240. King J, Unterkofler K, Teschl G, Teschl S, Mochalski P, et al. (2012) A modeling-based evaluation of isothermal rebreathing for breath gas analyses of highly soluble volatile organic compounds. *J Breath Res* 6: 016005.
241. King J, Unterkofler K, Teschl G, Teschl S, Koc H, et al. (2011) A mathematical model for breath gas analysis of volatile organic compounds with special emphasis on acetone. *J Math Biol* 63: 959–9.
242. Koc H, King J, Teschl G, Unterkofler K, Teschl S, et al. (2011) The role of mathematical modeling in VOC analysis using isoprene as a prototypic example. *J Breath Res* 5: 037102.
243. Martinez-Lozano P, Zingaro L, Finiguerra A, Cristoni S (2011) Secondary electrospray ionization-mass spectrometry: breath study on a control group. *J Breath Res* 5: 016002.
244. Wang C, Sahay P (2009) Breath analysis using laser spectroscopic techniques: breath biomarkers, spectral fingerprints, and detection limits. *Sensors* 9: 8230–62.
245. Bos LDJ, Wang Y, Weda H, Nijssen TM, Janssen AP, et al. (2014) A simple breath sampling method in intubated and mechanically ventilated critically ill patients. *Resp Physiol Neurobiol* 191: 67–74.
246. Ruzsanyi V, Fischer L, Herbig J, Ager C, Amann A (2013) Multi-capillary-column proton-transfer-reaction time-of-flight mass spectrometry. *J Chromatog A* 1316: 112–8.
247. Španěl P, Smith D (2011) Progress in SIFT-MS: Breath analysis and other applications. *Mass Spectrom Rev* 30: 236–67.

248. Santonico M, Pennazza G, Capuano R, Falconi C, Vink TJ, et al. (2012) Electronic noses calibration procedure in the context of a multicentre medical study. *Sens Actuat B* 173: 555–61.
249. Valera JL, Togores B, Cosio BG (2012) Use of the electronic nose for diagnosing respiratory diseases. *Arch Bronconeumol* 48: 187–8.
250. Pennazza G, Santonico M, Agrò AF (2013) Narrowing the gap between breathprinting and disease diagnosis, a sensor perspective. *Sens Actuat B* 179: 270–5.

Submit your manuscript to Annex Publishers and benefit from:

- ▶ Convenient online submission
- ▶ Rigorous peer review
- ▶ Immediate publication on acceptance
- ▶ Open access: articles freely available online
- ▶ High visibility within the field
- ▶ Better discount for your subsequent articles

Submit your manuscript at
<http://www.annexpublishers.com/paper-submission.php>