

Emerging Coxsackievirus A6 Causing Hand-Foot-and-Mouth Disease in Children in Gabon

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Abstract

Hand-Foot-and-Mouth Disease (HFMD) is an epidemic childhood illness caused by enteroviruses including enterovirus A71 (EV-A71), coxsackievirus A16 (CV-A16) and coxsackievirus A6 (CV-A6). This disease mainly affects children under 5 years of age, causing typical skin rash such as papulovesicular rash on the palms, feet, and mouth. However, atypical clinical signs may include eruption in others anatomic sites, palmar and plantar desquamation and onychomadesis. Severe complications including encephalitis, myocarditis or meningitis can occur. Four children with HFMD visited the "Centre International de Recherches Médicales de Franceville" (CIRMF) in March 2018 in the south-east of Gabon. The aim of this study was to report clinical data, described virological investigations leading to identify the pathogen responsible for these cases of HFMD. Throat and nasal swabs were collected. First, a Real Time Polymerase Chain Reaction (PCR) was performed for the diagnosis of enteroviruses. Then, two conventional PCR for genotype identification and phylogenetic analysis were performed. Coxsackievirus A6 was detected in the four children. Phylogenetic analysis of the sequence of the VP1 gene showed it belonged to subgenotype D4 of Coxsackievirus A6. Thus, the health centers of the city were informed in order to set up a surveillance network of this disease. This is the first description of HFMD in Gabon showed the circulation of CV-A6. These preliminary data which highlights the circulation of Coxsackievirus A6 in children under five years old emphasize the importance of establishing surveillance for HFMD in Gabon.

Keywords: Hand-Foot-and-Mouth Disease; Coxsackievirus A6; Children; First Description in Gabon

List of abbreviations: CIRMF: Centre International de Recherches Médicales de Franceville; CV: Coxsackievirus; HFMD: Hand-Foot-and-Mouth Disease; EV: Enterovirus

Introduction

Hand-Foot-and-Mouth Disease (HFMD) is a common epidemic childhood illness caused by viral pathogens within the order *Picornaviridales*, in the family *Picornaviridae*, in the genus *Enterovirus* and the species A. The most common viruses causing outbreaks of HFMD are enterovirus A71 (EV-A71), coxsackievirus A16 (CV-A16) and coxsackievirus A6 (CV-A6) which has emerged in Finland and then worldwide since 2008 [1-5]. The characteristics of this disease defined by the World Health Organization are fever > 38 °C in children < 5 years of age, typical skin rash (papulovesicular rash on the palms or soles of the feet, or both, buttocks, knees, or elbows) with or without mouth ulcers [6]. This disease is transmitted by sneezing, by direct or indirect contact with saliva, secretions or feces. Usually, this contagious disease is mild and self-limiting in one week. However, atypical clinical signs may include eruption in others anatomic sites [6]. Severe complications including encephalitis, myocarditis or meningitis have been described, less frequently in association with HFMD [7]. In Europe, a study reported an outbreak of

HFMD in 2014 and 2015 in France, during which the most frequent serotype was CV-A6. Moreover, Coxsackievirus A6 infection was sometimes responsible for more than half the atypical form [4,8]. This infection can be benign and completely resolved in about ten days. However, studies described palmar and plantar desquamation and onychomadesis several weeks after the disease [7,9-11]. Furthermore, neurological and systemic complications such as encephalitis can occur after an EV-A71 infection [7,12]. Typical and atypical forms and the severity of the disease could be correlated with the enterovirus serotype [4]. An enterovirus EV-A71 vaccine is available in order to prevent severe HFMD [13,14]. In several countries such as China surveillance was established to evaluate the prevalence of this illness and the efficacy of the vaccine against EV-A71. It showed the necessity to develop a multivalent HFMD vaccine including the CV-A16 and emerging CV-A6 pathogen [2,13,14].

Few data on epidemics or cases of HFMD in Central Africa are available. In Gabon this disease associated with enteroviruses is undocumented. Four cases of HFMD from neighborhood children occurred in Franceville in 2018. It is the first clinical description of HFMD and the first molecular characterization of Coxsackievirus A6.

Methods

Sampling and Nucleic Acid Extraction

Throat and nasal flocced swabs “553C” (Copan, Diagnostic) were collected, placed in dry tubes and diagnosed for enteroviruses at the CIRMF. All parents gave oral informed consent for the tests. The CIRMF is authorized to establish an epidemiological surveillance and a molecular diagnosis of the diseases circulating in Gabon. Ribonucleic acid (RNA) was extracted from swabs with the QIAamp Viral RNA Mini Kit (Qiagen) according to the manufacturer’s instruction after pretreatment in saline (0.9%). The elution volume of RNA was 100µl.

Amplification

A Real Time Polymerase Chain Reaction (PCR) was performed using primers for amplification of 5’ untranslated region (UTR) of all enteroviruses with the kit SuperScript III One-Step RT-PCR Platinum Taq Invitrogen [15]. Each 25µl reaction mixture contained 5µl of 5X One Step RT-PCR buffer containing 12.5 mM magnesium chloride, 400 µM each deoxynucleoside triphosphate, 40 ng of bovine serum albumin per µl, 0.4 µM primers, 0.2 µM probe and 5 µl of eluted RNA. The PCR was performed using the forward primer 5’-ACATGGTGTGAAGAGTCTATTGAGCT-3’, the reverse primer 5’-CCAAAGTAGTCCGTTCCGC-3’ and the probe 5’-ATTAGCCGCATTCAGGGGCCGGA-3’ labeled at the 5’ ends with the FAM quencher and at the 3’ ends with the Black Hole Quencher 1. For genotypic identification a conventional nested PCR was performed using outer forward primer (224-VP3), outer reverse primer (222-VP1), inner forward primer (AN89-VP1) and inner reverse primer (AN88-VP1) targeted at the 357 pb VP1 partial sequence gene [16]. The Kit SuperScript III One-Step RT-PCR Platinum Taq (Invitrogen) was used for the first round. Each 25-µl reaction mixture contained 1X buffer, 0.1 mM of magnesium sulfate, 40 ng/µl of bovine serum albumin, 0.4 µM of primers, 1 µl of Taq and 5µl of eluted RNA. The Kit DNA+Taq Polymerase (Invitrogen) was used for the second round with the same final volume and concentration of reagent, 1 µl of Platinum Taq (Invitrogen) and 2µl of amplification product of the first PCR. The first round was done with the following conditions: 30 min at 42 °C, 3 sec at 95 °C, followed by 10 cycles at 95 °C for 20 sec, a decrease of one degree per cycle from 52 °C to 43 °C and 72 °C for 1 min, followed by 40 cycles at 95 °C for 20 sec, 42 °C for 30 sec and 72 °C for 1 min, and a final elongation at 72 °C for 10 min. The second round of PCR was run for 3 min at 95 °C, followed by 45 cycles at 95 °C for 15 sec, 42 °C for 20 sec and 72 °C for 30 sec, followed by 72 °C for 10 min.

Genotype Identification

A second conventional PCR was performed using two rounds with the same kit and concentration of reagent as previously and specific nested primers based on the CV-A6 Gdula strain of VP1 to determine the genotype (outer sense 5’-GARGCTAACATYATAGCTCTTGGAGC-3’, outer antisense 5’-CCYTCATARTCHGTGGTGGTTATGCT-3’, inner sense 5’-GACACYGAYGARATYCAACAAACAGC-3’, inner antisense 5’-CGRTCRTTGCAGTGTTWGTATTGT-3’ [3]. The first round was done with the following conditions: 30 min at 50 °C, 10 min at 95 °C, followed by 40 cycles at 94 °C for 40 sec, 53 °C for 40 sec and 72 °C for 1 min, and a final elongation at 72 °C for 10 min. The second round of PCR was run for 10 min at 95 °C, followed by 40 cycles at 94 °C for 40 sec, 53 °C for 40 sec and 72 °C for 1 min, and a final elongation at 72 °C for 10 min. These generated amplicons of 891 pb were purified and sequenced by the method of Sanger with a 3500 Genetic Analyzer (Applied Biosystems) using a BigDye Terminator Ready Reaction Cycle sequencing kit. Phylogenetic analyses were performed using a multiple sequence alignment of the sequence obtained and a selection of reference strains from the GenBank database using the Basic Local Alignment Search Tool (BLASTn). Phylogenetic relationship was determined with MEGA 4.0 software. Multiple sequence alignments were created using ClustalX (version 1.81). The phylogenetic tree was built using the maximum-likelihood method with the PhyML algorithm [17] and drawn using FigTree v.1.4.0.

Results

Clinical Description

In March 2018 three children aged three years old, from the same neighborhood and school, and, a sibling aged 13 months visited the medical office of the *Centre International de Recherches Médicales de Franceville* (CIRMF) showing symptoms of hand-foot-

and-mouth disease (Table 1). All four children had a rash on the palms, mouth, feet and knees and fever (38 °C). The infant had an additional atypical form and showed a vesicle rash on the upper limbs and shoulders. One of three-years-old children who had an additional eruption on the elbows and a runny nose presented a palmar and plantar desquamation and onychomadesis one month after the symptoms disappeared (Table 1).

	Case 1	Case 2	Case 3	Case 4
Age	3-years-old	3-years-old	13-months-old	3-years-old
Sex	Male	Male	Male	Male
Week of the onset symptom	Week 11 (2018)	Week 11 (2018)	Week 11 (2018)	Week 12 (2018)
Epidemiological linkage	Same school and same class as cases 2 and 4	Same school and same class as cases 1 and 4	Brother of the case 2	Same school and same class as cases 1 and 2
Fever	38 °C	38 °C	38.8 °C	38 °C
Cutaneous symptoms	Vesicular skin rash on mouth, hands, elbows, knees and feet	Vesicular skin rash on mouth and feet	Vesicular skin rash on mouth, hands, arms, shoulders, knees and feet	Vesicular skin on mouth, hands and feet
Others symptoms	Palmar and plantar desquamation and onychomadesis after one month	No other symptoms	No other symptoms	Runny nose

Table 1: Clinical data of the four cases of HFMD

Virological Investigation

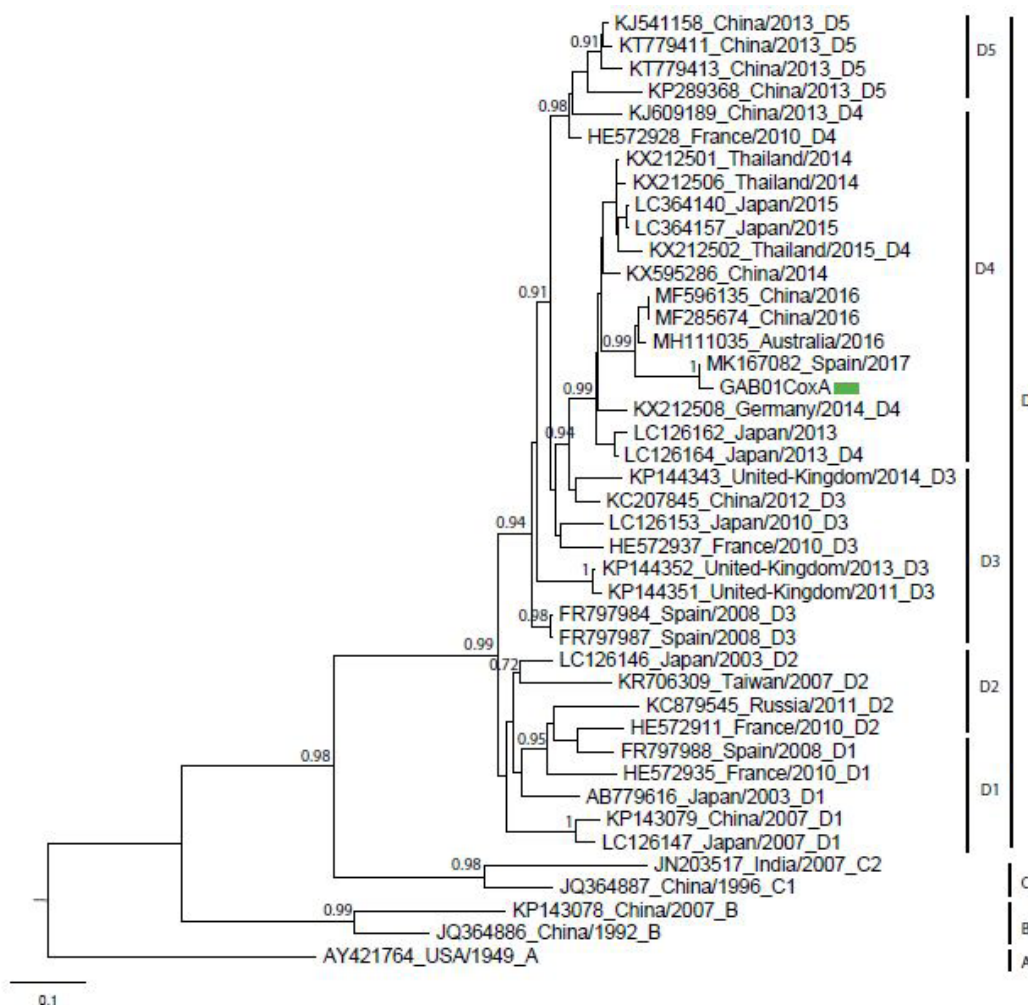


Figure 1: Molecular characterization of coxsackievirus strains; Phylogenetic tree for nucleotide sequences of Coxsackievirus A6 strains VP1 gene. The green rectangle indicates the sequence of Gabon (GAB01CoxA)

Enteroviruses were detected in the four samples. The child with a palmar and plantar desquamation and onychomadesis had the highest viral load. Four identical sequences were obtained from the samples of the four children. The sequence (807 pb) from the four isolates, named GAB01CoxA is available in the DDBJ/EMBL/GenBank database under accession numbers MK433298. A phylogenetic analysis based on 807 nucleotides sequence of the VP1 gene was performed using the sequence GAB01CoxA, 41 sequences of Coxsackievirus A6, from the GenBank database (Figure 1). We used the sequence of the Gdula strain prototype (genotype A) isolated in the USA in 1949 and genotype sequences B, C and D. The Gabonese strain displayed 99% to 94% identity at the nucleotide level with GenBank strains of genotypes D4 circulating in Europe and Asia between 2014 and 2017 (Figure 1). The Catalonia strain circulated in Spain in 2017 (Accession no. MK167082) have 99% identity with the Gabonese strain.

Discussion

We reported four cases of HFMD among neighbor children under five years old. These cases occurred in Franceville, a town in the south of Gabon, in March 2018. Molecular diagnostics showed that an enterovirus infected the patients. More specific molecular investigations showed that the children were infected by Coxsackievirus A6. These results corroborated those of several studies in Europe and Asia that reported the circulation of Coxsackievirus A6 causing HFMD [2,4]. One of the children showed an atypical form with a vesicle rash on the upper limbs and shoulders. Another had palmar and plantar desquamation and onychomadesis one month after contracting HFMD. These two atypical dermatologic presentations correlate studies which reported that CV-A6 was more frequently associated with unusual forms compared to other enterovirus serotypes [4,7,18]. However, the association between CV-A6 infection and onychomadesis is still widely debated in the literature [7]. Alignment was made with the four sequences (807 pb) obtained showing that they were strictly identical. The strain of this report (isolate GAB01CoxA, accession number MK433298) clustered with Coxsackievirus A6 and belonged to subgenotype D4. The homology (99%) between the Gabonese strain and this Spanish strain suggests that the Gabonese strain was recently imported in Gabon. However, the low number of cases and the lack of data concerning the circulation of CVA6 in Gabon, does not exclude their circulation and does not allow us to conclude that this strain is endemic or imported. An epidemiological survey was carried out. Patients did not travel outside Gabon during two months before onset. However, there are many exchanges and travels between Franceville, Europe and China for tourist and especially professional reasons. Since 2010, it would seem that most of the outbreaks of CV-A6 worldwide were due to genotype D3 [19]. However, these past few years, subgenotypes D4 and D5 emerged in Europe and Asia [3]. Subgenotype D4 was described in Finland and Germany, in 2008 and 2014 respectively. In China, several subgenotypes including D4 circulated in 2010, 2013 and 2015. Despite the recent emergence of D4, publications described a lower circulation [3,20]. In Central Africa and Gabon, little data on HFMD were available. No studies mentioned the circulation of CV-A6 and the genotypes circulating in Gabon. It may be due to the fact that this disease has not been described much because the clinical symptoms resemble those of a mild allergy. Indeed, other children from the same school and city appeared to have had the disease in the previous weeks even though no diagnosis was made. Thus, the health centers of the city were informed in order to set up a surveillance network of this disease. The number of children affected by HFMD in the same time period could have been underestimated. In France, it was showed that there was a gap existed in knowledge of the epidemiology and clinical impact of HFMD or herpangina because virological diagnosis wasn't performed. Therefore, the National Reference Laboratory for Enterovirus and Parechovirus set up surveillance in different regions of France from April 2014 to March 2015 reported that CV-A6 dominated the two epidemic occurred during this period [4]. These four cases of HFMD, described for the first time in Gabon suggested that CV-A6 circulated in the country and probably that ambulatory pediatrics cases could not have been investigated for virological diagnosis such as reported in France. A surveillance for HFMD would allow a better understand of the burden of HFMD, the epidemiology of enteroviruses such as CV-A6 and their serotypes. Previous studies have described that enteroviruses involved in HFMD can induce neurological complications [7,12,20]. Furthermore, a surveillance network of neurological syndrome has already been established in Franceville and Libreville, the capital of Gabon. So, first, we will include the diagnosis of Coxsackievirus in the surveillance system of neurological syndrome. Simultaneously, we set up a surveillance system of HFMD in the two main hospitals of Franceville. We informed the medical staff by describing the WHO case definition and the samples will sent to the CIRMF with clinical and epidemiological data for viral diagnosis.

Conclusion

These preliminary data which highlights the circulation of Coxsackievirus A6 in children under five years old emphasize the importance of establishing surveillance for HFMD in Gabon. We started the establishment of this network including also the viral diagnosis of neurological disorders.

Acknowledgment

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Conflicts of Interest

The authors declare no conflicts of interest.

Consent to Participate

The “Centre International de Recherches Médicales de Franceville” (CIRMF) is authorized to establish an epidemiological surveillance and a molecular diagnosis of the diseases circulating in Gabon. All parents of the four children gave verbal consent. The work described has been carried out in accordance with The Code of Ethics of the World Medical Association (Declaration of Helsinki).

References

1. Anh NT, Nhu LNT, Van HMT, Hong NTT, Thanh TT, et al. (2018) Emerging Coxsackievirus A6 Causing Hand, Foot and Mouth Disease, Vietnam. *Emerg Infect Dis* 24: 654-62.
2. Bian L, Wang Y, Yao X, Mao Q, Xu M (2015) Coxsackievirus A6: a new emerging pathogen causing hand, foot and mouth disease outbreaks worldwide. *Expert Rev Anti Infect Ther* 13: 1061-71.
3. He S, Chen M, Wu W, Yan Q, Zhuo Z (2018) An emerging and expanding clade accounts for the persistent outbreak of Coxsackievirus A6-associated hand, foot, and mouth disease in China since 2013. *Virology* 518: 328-34.
4. Mirand A, Le Sage FV, Pereira B, Cohen R, Levy C (2016) Ambulatory Pediatric Surveillance of Hand, Foot and Mouth Disease as Signal of an Outbreak of Coxsackievirus A6 Infections, France, 2014-2015. *Emerg Infect Dis* 22: 1884-93.
5. Osterback R, Vuorinen T, Linna M, Susi P, Hyypia T (2009) Coxsackievirus A6 and hand, foot, and mouth disease, Finland. *Emerg Infect Dis* 15: 1485-8.
6. World Health Organization (2011) A Guide to Clinical Management and Public Health Response for Hand, Foot and Mouth Disease (HFMD), Geneva, Switzerland.
7. Mammas IN, Theodoridou M, Kramvis A, Thiagarajan P, Gardner S, et al. (2017) Paediatric Virology: A rapidly increasing educational challenge. *Exp Ther Med* 13: 364-77.
8. Folster-Holst R (2018) Classical Hand, Foot and Mouth Disease Replaced by Atypical Hand, Foot and Mouth Disease. *Acta Derm Venereol* 98: 303.
9. Bernier V, Labreze C, Bury F, Taieb A (2001) Nail matrix arrest in the course of hand, foot and mouth disease. *Eur J Pediatr* 160: 649-51.
10. Clementz GC, Mancini AJ (2000) Nail matrix arrest following hand-foot-mouth disease: a report of five children. *Pediatr Dermatol* 17: 7-11.
11. Kaminska K, Martinetti G, Lucchini R, Kaya G, Mainetti C, et al. (2013) Coxsackievirus A6 and Hand, Foot and Mouth Disease: Three Case Reports of Familial Child-to-Immunocompetent Adult Transmission and a Literature Review. *Case Rep Dermatol* 5: 203-9.
12. Solomon T, Lewthwaite P, Perera D, Cardoso MJ, McMinn P, et al. (2010) Virology, epidemiology, pathogenesis, and control of enterovirus 71. *Lancet Infect Dis* 10: 778-90.
13. Fang CY, Liu CC (2018) Recent development of enterovirus A vaccine candidates for the prevention of hand, foot, and mouth disease. *Expert Rev Vaccines* 17: 819-31.
14. Li J, Pan H, Wang X, Zhu Q, Ge Y, et al. (2018) Epidemiological surveillance of hand, foot and mouth disease in Shanghai in 2014-2016, prior to the introduction of the enterovirus 71 vaccine. *Emerg Microbes Infect* 7: 37.
15. Dierssen U, Rehren F, Henke-Gendo C, Harste G, Heim A (2008) Rapid routine detection of enterovirus RNA in cerebrospinal fluid by a one-step real-time RT-PCR assay. *J Clin Virol* 42: 58-64.
16. Nix WA, Oberste MS, Pallansch MA (2006) Sensitive, seminested PCR amplification of VP1 sequences for direct identification of all enterovirus serotypes from original clinical specimens. *J Clin Microbiol* 44: 2698-704.
17. Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W (2010) New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. *Syst Biol* 59: 307-21.
18. Puenpa J, Vongpunsawad S, Osterback R, Waris M, Eriksson E, et al. (2016) Molecular epidemiology and the evolution of human coxsackievirus A6. *J Gen Virol* 97: 3225-31.
19. Song Y, Zhang Y, Ji T, Gu X, Yang Q, et al. (2017) Persistent circulation of Coxsackievirus A6 of genotype D3 in mainland of China between 2008 and 2015. *Sci Rep* 7: 5491.
20. Lau SKP, Zhao PSH, Sridhar S, Yip CCY, Aw-Yong KL, et al. (2018) Molecular epidemiology of coxsackievirus A6 circulating in Hong Kong reveals common neurological manifestations and emergence of novel recombinant groups. *J Clin Virol* 108: 43-9.

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