

Toxic Effect of Titanium (TiO₂) on Wistar Rat (*Rattus Norvegicus*) Injected by Intravenously

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Abstract

Titanium dioxide (TiO₂) (<100 nm) has been widely used in the production of paints, paper, and plastics, as well as in food additives and colorants. It has been classified as biologically inert in both humans and animals. Here we studied the toxicity of TiO₂ on Wistar rat. Main exposure area of NPs is caudal vein of Wistar rat. In the field of nanomedicine, intravenous injection can deliver TiO₂ nanoparticulate carriers directly into the human body. Upon intravenous exposure, TiO₂ NPs can induce pathological lesions of the liver, spleen, kidneys, and brain. We have also shown here that most of these effects may be due to the use of very high doses of TiO₂ NPs. Ultrafine size and morphological properties of NPs was determined by TEM, SEM and X ray diffraction where the size of NPs were 5-20nm, found morphologically as aggregate form. Due to high dose of NPs increase of body weight was reduced but the coefficient of liver, kidney and spleen were significantly increased but no change in brain. Reactive Oxygen Species (ROS) increase due to high doses which damaged DNA. Serum protein and catalase enzyme activity also increased with increasing NPs concentration but SOD reduced.

Keywords: Wistar Rat; Titanium; *Rattus Norvegicus*

List of Abbreviations: NPs-Nanoparticles; TEM-Transmission Electron Microscopy; SEM-Scanning Microscopy; ROS-Reactive Oxygen Species; SOD-Super Oxide Dismutase

Introduction

Nanotechnology involves the design, production, characterization and application of nanoparticles whose dimensions are less than 100nm. Toxicity of nanoparticle depends on particle size, particle composition, and specific cell type. Nanoparticles can be translocated to the sub epithelial space to a greater extent than larger particles [1]. The interactions of particles with cell membranes results in the generation of reactive oxygen species (ROS) [2], and the generated oxidative stress may cause a breakdown of membrane lipids, an imbalance of intracellular calcium homeostasis, and DNA breakage [3]. Ultrafine particles are known to have greater adverse health effects than larger particles [4] because of their extremely high surface areas and the ability to deposit in the alveoli. In October 2011 the European Union defined nonmaterial as a natural, incidental or manufactured material containing particles, in an unbound state or as an aggregate or agglomerate; where 50% or more of the particles exhibited, one or more external dimensions in the size range 1–100 nm [5]. Others have defined NPs as objects with at least one of their three dimensions in the range of 1–100 nm [6,7]. NPs generally possess dramatically different physicochemical properties compared to fine particles (FPs) of the same composition. The smaller size of NPs ensures that a large portion of atoms will be on the particle surface. Traditionally, TiO₂ NPs have been considered as poorly soluble, low toxicity particles [8,9]. Due to this reason, they have been traditionally used as a “negative control” in many *in vitro* and *in vivo* particle toxicological studies [10]. Oberdorster et al. reported that TiO₂ NPs (21 nm) caused a greater pulmonary inflammatory response than TiO₂ at same mass burden, with greater amounts of TiO₂ NPs entering the alveolar interstitium in the lungs [11]. Sager et al. [2008] have reported similar results after intra-tracheal instillation of well-dispersed suspensions of TiO₂ NPs (80/20 anatase/rutile; 21 nm, P-25) and TiO₂ FPs (100% rutile; 1 μm) in rats. On an equal mass burden, nano TiO₂ was 40 fold more potent in inducing lung inflammation and damage at 1 and 42 days post-exposure than fine TiO₂ [12]. However, respective potencies were not significantly different when dose was expressed on the basis of total surface area of particles delivered to the lung. Wide application of TiO₂ NPs confers substantial potential for human exposure and environmental release, which inevitably allows for a potential health risk to humans, livestock. This paper will focus mainly on current knowledge concerning the toxicology of TiO₂ NPs. Studies done with mixtures of TiO₂ NPs and studies that have focused on wistar rat and the environment will not be discussed in this review. Even though the nanoparticle (NP) size has recently been defined as <100 nm, we have also included some studies that have defined particle sizes that are >100 nm as NPs.

Material and Methods

Chemicals and animal

Hydroxypropyl methyl cellulose (HPMC), phosphate buffer saline (PBS), ethanol, phosphoric acid, sodium acetate, ferrous sulphate

hate, epinephrine, hydrochloric acid sodium carbonate, H_2O_2 and Animal (purchased from national Institute of Nutrition (NIN) Hyderabad, India).

Nanoparticle solution

TiO_2 was purchased from Sigma Aldrich Pvt Ltd Kushaiguda, Kamla Nagar, besides Mc Donalds, Hyderabad, India. It was sonicated 35 KHZ for 20 minutes because particles got agglomerated. Ultra sonication disrupts agglomerated form of nanoparticles come under nanometer size.

Characterization of TiO_2 nanoparticles

TiO_2 nanoparticles (TNPs) were dissolved in 0.5% HPMC (Hydroxypropyl methyl cellulose) and ultrasonicated for 10 min to make a homogeneous suspension. Average particle size, morphology and size distribution of TNP were examined by a JEOL-JEM-2100F Transmission electron microscope (TEM) operating at 200 kV. The aqueous suspension was drop casted on a carbon coated copper grid, and the grid was air dried at room temperature (25 °C) before loading into microscope. The TNPs were further characterized by X-ray diffraction (XRD). The XRD patterns (Bragg peaks) were recorded with a PANalytical X'pert PRO diffractometer using a solid state detector with a monochromatized Cu $\alpha 1$ ($\lambda_{Cu} = 1.54060 \text{ \AA}$) radiation source at 45 kV. For morphological analysis it was homogeneously spreaded over the glass plate and air-dried in vacuum and SEM (Scanning Electron Microscopy) image were observed with ZEISS EVO40, SEM, at 10 Kv.

Treatment of animal with TiO_2 nanoparticles

Six weeks old (225-250gm) male Wistar rats were subjected for the in-vivo toxicity study of nano anatase TiO_2 . Animals were divided into three different groups, with six animals in each group. Each animal was injected intravenously (caudal vein) with different doses of NPs. Group I was given 5 mg/ kg body weight (BW) of animal, group II was injected with 50 mg/kg BW and Group III was injected with 0.5% HPMC, treated as control. TSNPs was injected at weekly interval for 30 days.

Blood Sampling

Animals were anesthetized with 0.3ml/250mg ketamine/xylazine and 1 ml blood was taken by retro-orbital veins through heparin coated capillaries and blood was collected in tubes containing 20 mg/ml EDTA. At the end of the experiment all animals were sacrificed at the same time after treatment and 5 ml blood was collected from heart. Serum was collected from blood and stored at -80 °C for different biochemical assays.

Coefficients of liver, kidney and spleen

Body weights of all animals were recorded before sacrifice. Animals were sacrificed by anesthetic ether and cervical dislocation. Different organs were dissected from the animals and were weighed separately. Coefficients of liver, kidney and spleen to body weight were calculated as per the ratio of tissue's weight (mg) to body weight (gm).

Reactive oxygen species assay

ROS assay was performed according to the method of Hayashi et al 1968, 5 μ l of serum sample was added to 140 μ l of pre warmed (40 °C) 0.1M sodium acetate buffer (pH 4.8) in one well of a 96-well (microtiter plate). Mixture of (100 μ l) of DEPPD (100 μ g/ml DEPPD was dissolved in 0.1M sodium acetate buffer, (pH 4.8) and ferrous sulfate [ferrous sulfate (4.37 μ M) was dissolved in 0.1M sodium acetate buffer, pH 4.8] was added in each well to initiate reaction. After 5 min incubation absorbance was measured using a Spectra Max M_2 spectrophotometer plate reader, at 505 nm. ROS levels in serum were calculated from the calibration curve of H_2O_2 and expressed as equivalent to levels of hydrogen peroxide (1 unit = 1.0 mg H_2O_2 /l). Calibration curve for the standard solution was obtained by calculating slopes from optical density graph.

Protein estimation

Total protein was assayed by Bradford method [13]. For each sample, 200 μ l of the serum was mixed in 800 μ l of Bradford reagent. Optical density (at 595 nm) was measured after 10 min of incubation in the dark. BSA was used as a standard protein. Optical density was obtained by spectrophotometer (Erba CHEM-5Plus v2).

Determination of Catalase activity

Serum catalase activity was determined by measuring the decrease in absorbance at 240nm due to the decomposition of H_2O_2 in a UV recording spectrophotometer. The reaction mixture (3 ml) contained 0.1 ml of serum in phosphate buffer (50 mM, pH 7.0) and 2.9 ml of 30 mM H_2O_2 in phosphate buffer pH 7.0. The specific activity of catalase was expressed as moles of H_2O_2 reduced per minute per mg protein.

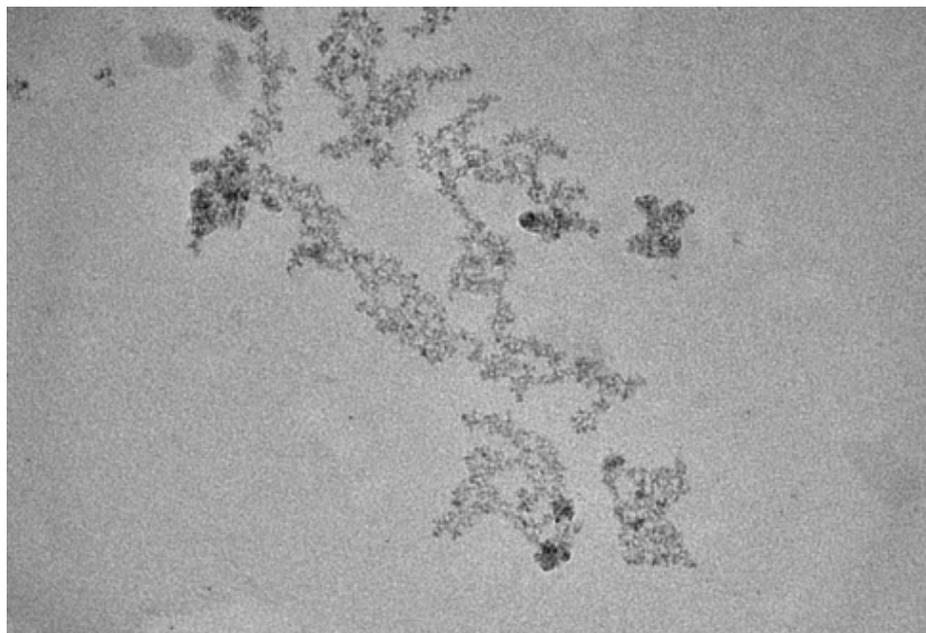
Determination of SOD activity

The reaction mixture (3 ml) contained 2.95 ml 0.05 M sodium carbonate buffer pH 10.2, 0.02 ml of serum and 0.03 ml of epinephrine in 0.005 N HCL was used to initiate the reaction. The reference cuvette contained 2.95 ml buffer, 0.03 ml of substrate (epinephrine) and 0.02 ml of water. Enzyme activity was calculated by measuring the change in absorbance at 480 nm for 5 min.

Results

Characterization of TNPs

The size of the TiO_2 nanoparticles (TNPs) was determined by transmission electron microscopy (TEM, JEOL-JEM-2100F). Figure 1 shows the typical TEM images of TNPs, from which it was observed that the TNPs had an average diameter of 5-20 nm. Clustering of NPs was also observed from the images. The wide-angle region of the X-ray diffraction (XRD, PANalytical X'pert PRO) patterns exhibited a high-intensity diffraction peak at $2\theta = 25.47^\circ$ and three additional peaks at $2\theta = 38.0^\circ$, 48.17° and 68.8° (Figure 2) with the respective size 15.3, 9.2, 7.3 and 10.4 nm. Size of TNPs calculated by XRD and TEM was $(5\pm 1-16\pm 1 \text{ nm})$ (Figure 2). The morphologies of the TNPs were observed via scanning electron microscopy (SEM, ZEISS EVO 40). As shown in Figure 3 nanoparticles were found in aggregate form, in this case the morphologies of TiO_2 nano could not clearly observed. The average diameter of the TiO_2 in SEM was $17\pm 2 \text{ nm}$, which was in agreement with TEM images.



Nano_005
Print Mag: 154000x @7.0 in

100 nm
HV=120.0kv
Direct Mag: 30000x
AIF-JNU

Figure 1: TEM image of TiO_2 NPs particle size is calculated with TEM and Range from 5-20 nm. Analysis was done at 200kv

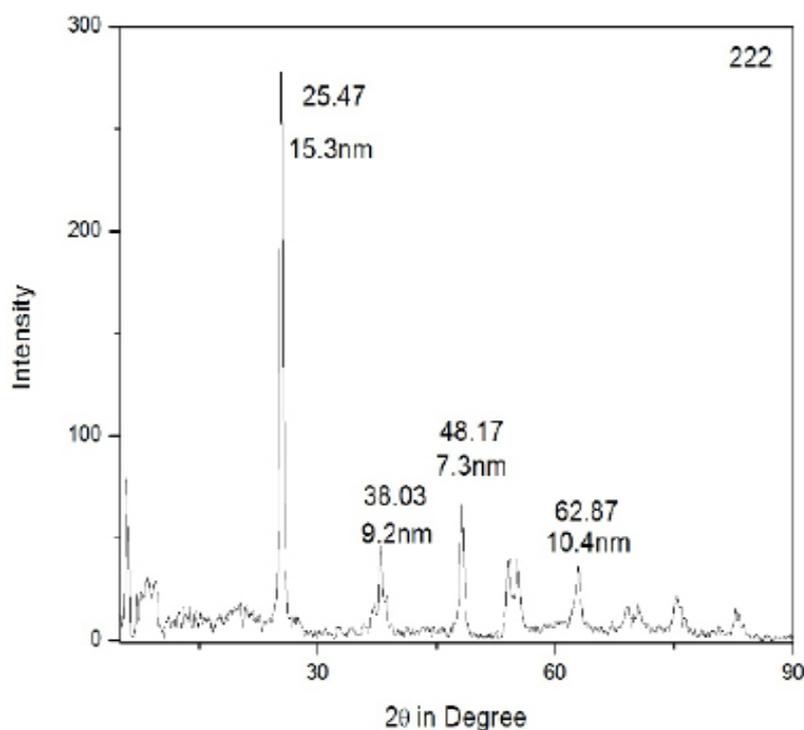


Figure 2: Characterization of TiO_2 Powder X-ray diffraction patterns of anatase TiO_2 with Miller indices (h k l) showing crystal family of planes for each diffraction peak

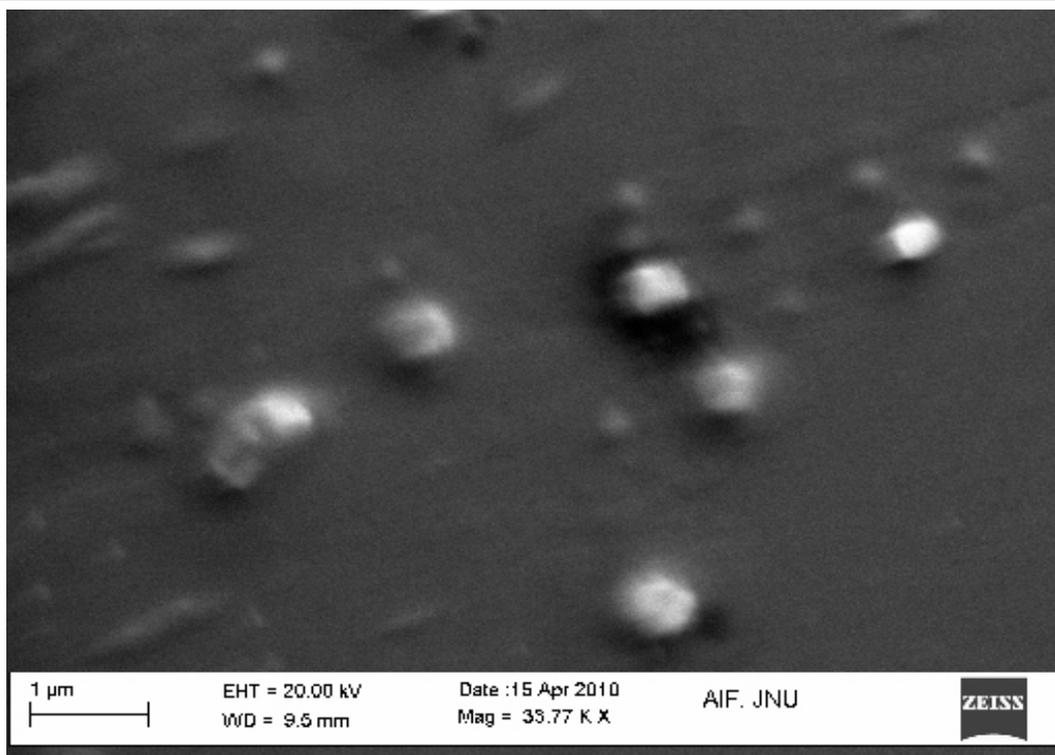


Figure 3: Morphology and microstructure characterization by SEM

Coefficient of organs

Coefficients of the liver, kidney, spleen and brain of animal treated with lower dose (5 mg/kg) of TNPs there were no significant changes except liver. Whereas animals treated with higher dose (50 mg/kg) of TNPs, the net increase in body weight were significantly reduced ($p < 0.05$) and coefficients of the liver, kidney and spleen were significantly ($p < 0.05$) increased, which suggested that higher dose of TNPs might cause the damage of the organs of rat. There were no significant ($p > 0.05$) changes observed in Coefficient of brain in all treatment groups (Figure 4).

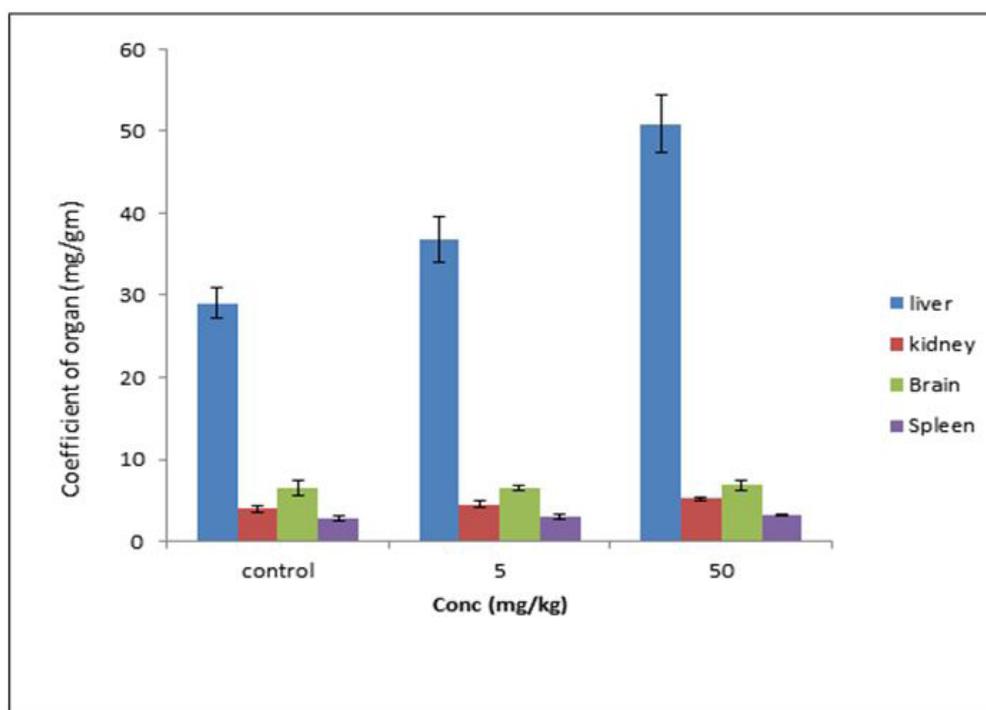


Figure 4: Coefficient of organs of rat treated with TiO_2 (liver, kidney, spleen, brain). Coefficient of organs is the ratio of weight of the organs (mg) to animal body weight (gm). Legend indicates the types of organs.

* denotes statically significant result at $\alpha=0.05$

** denotes statically significant result at $\alpha=0.01$.

Statistical analysis was performed with a two-sample t-test, unknown and unequal variances, comparing each sample group to the related control group.

Oxidative stress measurement

Oxidative stress is a key factor in the genotoxic studies. Oxidative stress was measured by DEPPD (N,N-diethyl-para-phenyldiamine) sulphate staining. It was found that level of total reactive oxygen species increased significantly ($p < 0.01$) with respect to the low to high dose of nano anatase TiO_2 (50 mg/kg BW) (Figure 5). Higher concentration of reactive oxygen species may be responsible for DNA damage and tissues injuries.

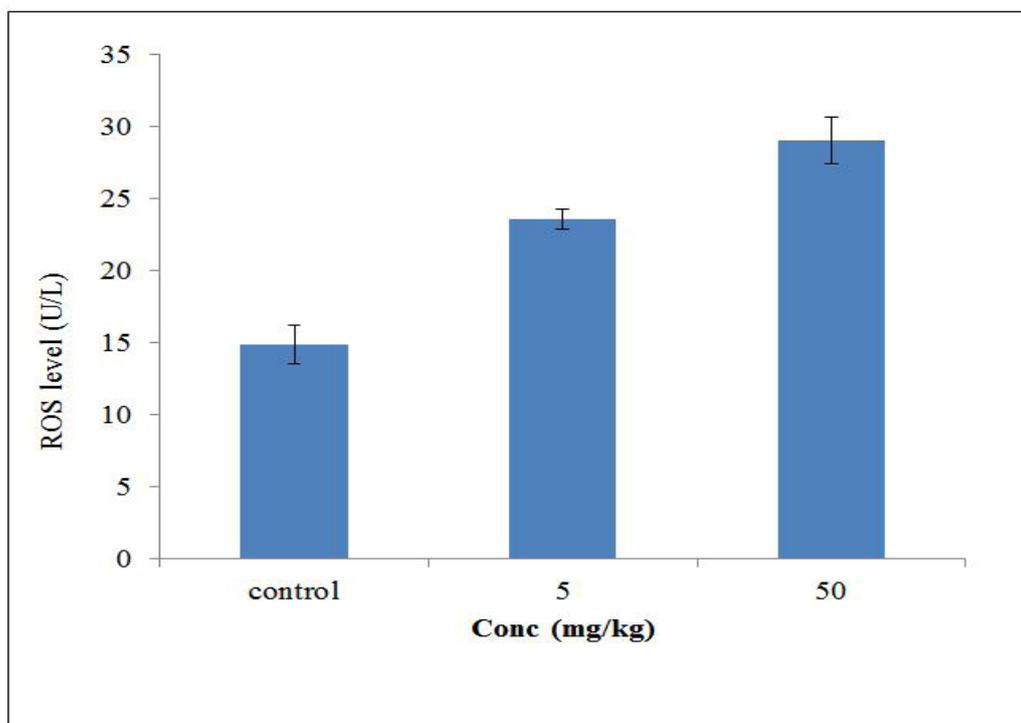


Figure 5: Reactive oxygen species measurement from wistar rat treated with various concentrations TiO_2 . Results illustrated mean and standard deviation. Error bar represent standard deviation. Statistical analysis was performed with a two-sample t-test, unknown and unequal variances, comparing each sample group to the related control group

Protein estimation, Catalase activity and SOD activity

There was a significant increase in the serum protein with 5 mg/kg and 50 mg/kg nano TiO_2 (Figure 6). Catalase activity was significantly decreased ($p < 0.05$) in rats treated with TiO_2 . However, this depressed activity was more significant in 50 mg/kg than 5 mg/kg (Figure 7). Results show a significant increase ($p < 0.05$) SOD activity of serum rats treated with 5 mg/kg and 50 mg/kg as compared to the untreated rats. This increase was more significant ($p < 0.05$) in 50 mg/kg treated animal than 5 mg/kg treated animal (Figure 8).

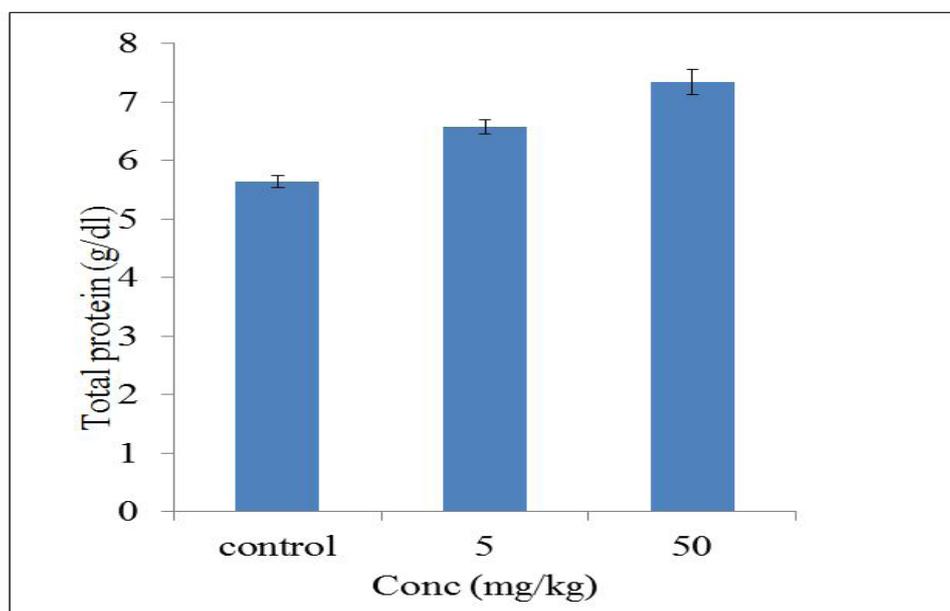


Figure 6: Serum concentration of wistar rat treated with varying concentration of nano TiO_2 . Results illustrate mean and standard deviation of total protein

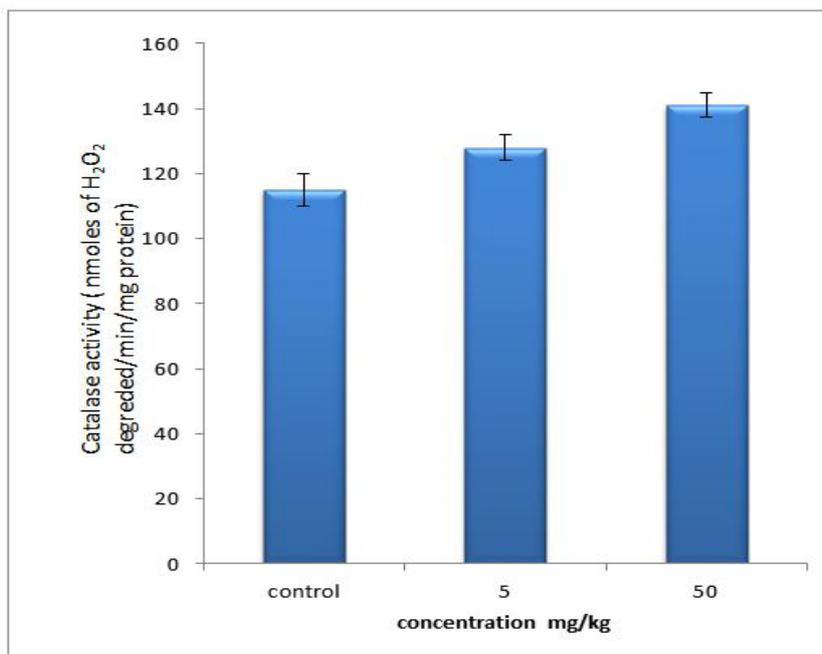


Figure 7: Catalase activity rats serum under control conditions and after the exposure of TiO₂ indicate significant differences ($p < 0.05$ and $p < 0.001$) from control rat

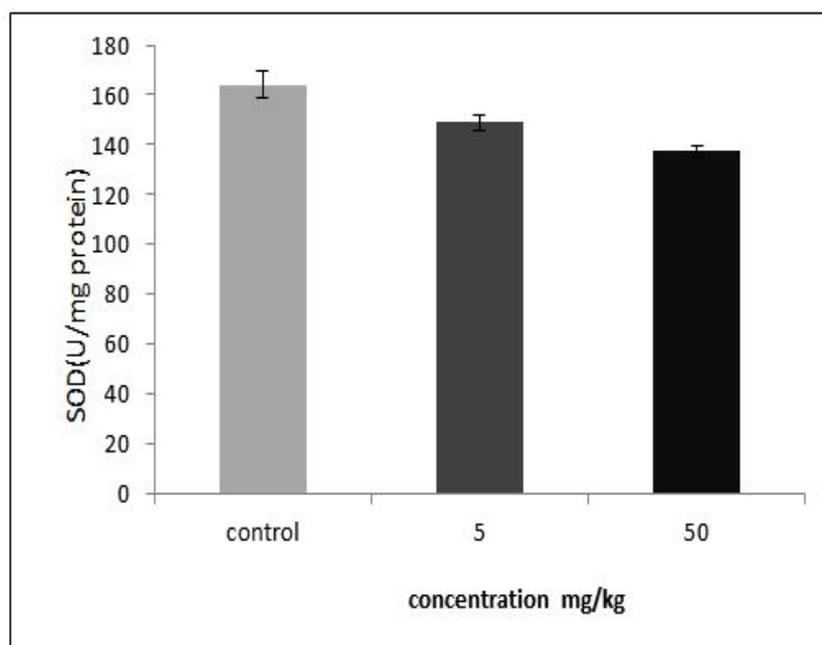


Figure 8: SOD of rats serum under control conditions and after the exposure of TiO₂ indicate significant differences ($p < 0.05$ and $p < 0.001$) from rats control conditions. It was decreased with increasing TiO₂ concentration

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